






Review

Reappraisal of the Concept of Accelerated Aging in Neurodegeneration and Beyond

Yauhen Statsenko ^{1,2,3,†} , Nik V. Kuznetsov ^{2,*,†}, Daria Morozova ² , Katsiaryna Liaonchyk ², Gillian Lylian Simiyu ¹, Darya Smetanina ¹, Aidar Kashapov ¹, Sarah Meribout ¹, Klaus Neidl-Van Gorkom ¹, Rifat Hamoudi ^{2,4,5} , Fatima Ismail ⁶, Suraiya Anjum Ansari ^{2,7} , Bright Starling Emerald ^{2,8}  and Milos Ljubisavljevic ^{2,9}

- ¹ Department of Radiology, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates; e.a.statsenko@uaeu.ac.ae (Y.S.); gl.simiyu@uaeu.ac.ae (G.L.S.); daryasm@uaeu.ac.ae (D.S.); a.kashapov@uaeu.ac.ae (A.K.); sarahmeribout@uaeu.ac.ae (S.M.); klausg@uaeu.ac.ae (K.N.-V.G.)
 - ² ASPIRE Precision Medicine Research Institute Abu Dhabi, United Arab Emirates University, Al Ain 27272, United Arab Emirates; darsmorozova@uaeu.ac.ae (D.M.); katherine.leonchik@gmail.com (K.L.); rhamoudi@sharjah.ac.ae (R.H.); sansari@uaeu.ac.ae (S.A.A.); bsemerald@uaeu.ac.ae (B.S.E.); milos@uaeu.ac.ae (M.L.)
 - ³ Big Data Analytic Center, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates
 - ⁴ Department of Clinical Sciences, College of Medicine, University of Sharjah, Sharjah 27272, United Arab Emirates
 - ⁵ Division of Surgery and Interventional Science, University College London, London NW3 2PS, UK
 - ⁶ Department of Pediatrics, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates; fatima.ismail@uaeu.ac.ae
 - ⁷ Department of Biochemistry and Molecular Biology, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates
 - ⁸ Department of Anatomy, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates
 - ⁹ Department of Physiology, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates
- * Correspondence: nikkuznetsov@inbox.com
† These authors contributed equally to this work.



Citation: Statsenko, Y.; Kuznetsov, N.V.; Morozova, D.; Liaonchyk, K.; Simiyu, G.L.; Smetanina, D.; Kashapov, A.; Meribout, S.; Gorkom, K.N.-V.; Hamoudi, R.; et al.

Reappraisal of the Concept of Accelerated Aging in Neurodegeneration and Beyond. *Cells* **2023**, *12*, 2451. <https://doi.org/10.3390/cells12202451>

Academic Editors: Pierre Jouanny, Sonia Hammami, Gérard Lizard and Anne Vejux

Received: 4 August 2023
Revised: 1 September 2023
Accepted: 6 September 2023
Published: 14 October 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Background: Genetic and epigenetic changes, oxidative stress and inflammation influence the rate of aging, which diseases, lifestyle and environmental factors can further accelerate. In accelerated aging (AA), the biological age exceeds the chronological age. Objective: The objective of this study is to reappraise the AA concept critically, considering its weaknesses and limitations. Methods: We reviewed more than 300 recent articles dealing with the physiology of brain aging and neurodegeneration pathophysiology. Results: (1) Application of the AA concept to individual organs outside the brain is challenging as organs of different systems age at different rates. (2) There is a need to consider the deceleration of aging due to the potential use of the individual structure–functional reserves. The latter can be restored by pharmacological and/or cognitive therapy, environment, etc. (3) The AA concept lacks both standardised terminology and methodology. (4) Changes in specific molecular biomarkers (MBM) reflect aging-related processes; however, numerous MBM candidates should be validated to consolidate the AA theory. (5) The exact nature of many potential causal factors, biological outcomes and interactions between the former and the latter remain largely unclear. Conclusions: Although AA is commonly recognised as a perspective theory, it still suffers from a number of gaps and limitations that assume the necessity for an updated AA concept.

Keywords: aging; accelerated aging; brain aging; neurodegeneration; epigenetics; biological clocks; molecular biomarkers; rejuvenation

1. Introduction

Aging is associated with structural and physiological changes that increase the risk of developing diseases and death [1–3]. Aging processes are influenced by different factors, including genetic mutations, epigenetic modifications, oxidative stress and inflammation [4], resulting in the accumulation of damage and dysfunction at all biological levels [5]. Biological age (BA), also called physiological age, can be defined as the current state of an individual as a biological system, characterized by a combination of detectable life time-dependent biological parameters (determination criteria), for example, by the current profile of genomic DNA methylation, the present status of aging-associated structures in the brain, etc.

Normal aging corresponds to all monitored aging-associated processes where BA equals the chronological age. Accelerated aging (AA) is observed when the BA exceeds the chronological age and, vice versa, decelerated aging is observed when the chronological age exceeds the BA [6–8]. AA shares common features with normal aging, but it is also characterised by specific processes such as protein aggregation and excitotoxicity [9–11]. Understanding the mechanisms of aging can open opportunities for targeted therapies to slow it [9].

AA is an area of active research with unresolved issues, including the unstandardised terminology [12] and understudied mechanisms [13] including neurodegeneration (ND) described either as a type of AA [14,15] or as its outcome [14,16–18]. The latter view argues that certain biomarkers (BM) are ND-specific and do not detect AA [17]. Different theories have been proposed to explain the pathogenesis of AA, including genetic theory [19], the multi-proteinopathies theory [20] and mitochondrial theory [21]. Genetic theory assumes the accumulation of DNA mutations and/or gene dysregulation in AA and has certain limitations [19,22]. It considers random DNA changes but ignores chromosomal, multifactorial and monogenic alterations [13,23,24]. The multi-proteinopathies theory is based on the accumulation/aggregation of misfolded proteins leading to cell dysfunction and causing age-related diseases [20,25]. The free radical theory considers the oxidative damage to DNA and proteins by reactive oxygen species (ROS) as the primary accelerator of aging [15,26–28]; however, it has the problem of segregating between the normal and abnormal levels of ROS [21,29]. However, these aging theories lack reliable diagnostic BM for early identification and prognostication [20,30].

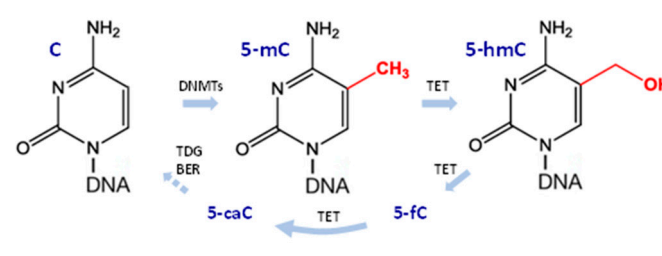
2. Biomolecular Aspects of Aging

Aging theories differ in their approaches to describing aging at cellular, supracellular and subcellular levels. ND aetiology can be approached with the neurocentric (NC) or neurovascular (NV) view. The latter refers to the dynamic multicellular structure called the neurovascular unit (NVU) that includes astrocytes, microglia, oligodendrocytes, precursor cells, excitatory and inhibitory neurons, endothelial cells and pericytes, and mediates the functional interactions between brain tissues per se and the blood vessels [31]. The NV hypothesis proposes that neural cells in the NVU and circulating immune cells secrete pro-inflammatory mediators, therefore, contributing to age-related neuroinflammation [32], cell degeneration [33,34] and endothelial impairment [34,35]. These changes disrupt molecular networks, induce damage to the blood–brain barrier [36,37] and lead to NVU dysfunction, a major cause of ND [38]. However, the exact role of NVU in ND remains unclear [39]. Accumulated evidences of the high complexity and molecular heterogeneity of the NVU network makes the search for associated BM difficult and requires whole genome studies, e.g., global transcriptome analysis followed by hierarchical data clustering [40] or single-cell/single-nucleus transcriptomics [41,42].

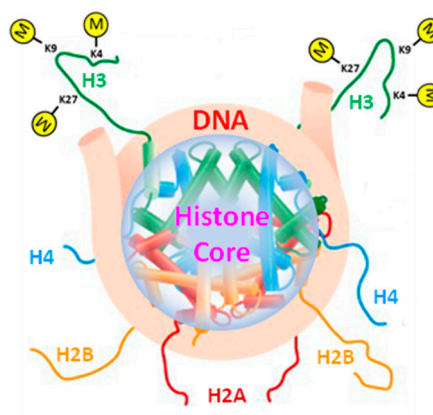
Molecular biomarkers (MBM) are biomolecules, their components, fragments or modifications with associated measurable parameters that serve as a tool to diagnose pathologies and monitor biological processes. MBM can be used to evaluate aging, particularly to estimate the rate of its progression [43]. Aging MBM include mRNA transcripts, proteins [44], the length of telomeres, serum markers of DNA damage [45], DNA methylation

profiles [46,47], histone modifications [48–59], differentially expressed genes [42,60], non-coding RNAs [61–64] and other biomolecules (Figure 1).

I. DNA nucleotide modification level



II. Histone modification level



III. Regulatory RNAs level

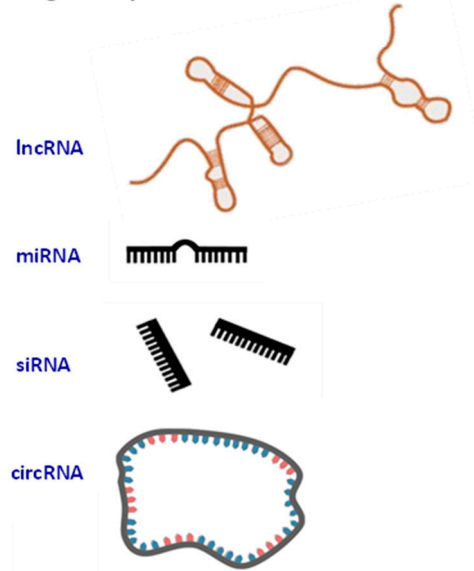


Figure 1. Examples of epigenetic changes and factors affected by accelerated aging at different levels that could serve as potential biomarkers of aging. (I) DNA nucleotide modification level: cytosine (C) can be converted to 5-methylcytosine (5-mC) and oxidised further to 5-hydroxymethylcytosine (5-hmC). (II) Histone modification level: Three examples of histone 3 (H3) lysine methylation marks are indicated. Lysine 4 (K4) H3K4me3 mark is associated with active genes, lysine 9 (K9) H3K9me3 is an established mark of transcriptionally repressed heterochromatin and lysine 27 (K27) H3K27me3 mark is linked to both transcriptional activation and repression. (III) Regulatory RNAs level: several types of ncRNAs, long non-coding RNA (lnc RNA), microRNA (miRNA), small interfering RNA (siRNA), circular RNA (circRNA), shown schematically.

Despite the large number of suggested MBM of ND, only a few of them have been validated. In most studies, the sample sizes have been too small to justify the accuracy and reproducibility of MBM data. For example, a recent study aimed at determining whether age affects different cell types in NVU resulted in the model discriminating Alzheimer's disease (AD) from healthy control (HC) samples, revealing 15 genes related to accelerated aging (AAG): *IGF1R*, *MXII*, *RB1*, *PPARA*, *NFE2L2*, *STAT5B*, *FOS*, *PRKCD*, *YWHAZ*, *HTT*, *MAPK9*, *HSPA9*, *SDHC*, *PRKDC* and *PDPK1* [42]. Of these genes, differential expression of *IGF1R*, *MXII*, *PPARA*, *YWHAZ* and *MAPK9* correlate with the progression of ND and may function as facilitators or inhibitors of AD. However, questions remain on the cell-specific roles of the discovered AAGs, and their contribution to AD pathogenesis and interactions in NVU. Moreover, the study cohort included only 11 AD patients and seven HC, which is insufficient for justifying AAGs as MBM in AD [42]. ND results from multiple structural changes at different genetic loci over a period of time [65,66]. AD represents 90% of ND cases, and the chances of it developing increase with changes in the 15 indicated genes predisposing to ND (NDG): *GBA1*, *APP*, *PSEN1*, *MAPT*, *GRN*, *SETX*, *SPAST*, *CSF1R*, *C9orf72* [67], *TET2* [68], *TBK1* [69], *TOMM40*, *APOC1* [70], *APOE* [70,71] and *TREM2* [72–78]. Surprisingly, there is no overlap between both sets of genes resulting from different studies: AAGs and NDGs. In dementia with Lewy bodies, multi-cognitive decline and corticobasal degeneration, the risk factors shown to be involved are the *APOE* e4 allele and the mutation spectrum for *TREM2* gene [78–87].

DNA methylation rate reflects the rate of aging. Approximately 1.5% of genomic DNA contains 5-methylcytosine (5-mC) that decreases during ontogenesis [88]. The level of 5-mC is found to be highest in embryos and it decreases gradually with age [89,90]. Global genomic DNA hypomethylation in aging proceeds along with hypermethylation of CpG islands (CGIs) in the mammalian genome where 60% of CGIs are associated with gene promoters and involved in the regulation of gene transcription [91]. Changes in DNA methylation patterns known as “epigenetic drift” are associated with aging across the entire lifespan [92].

Age-predictive models demonstrate gradual linear changes in the DNA methylation profile in normal aging; however, environmental or genetic risk factors accelerate aging [93]. In monozygotic twins, the divergence of the methylome increases at different rates [94]. Furthermore, the DNA methylation profile was proposed as mechanisms of the epigenetic clock [95–97] by analogy with the biological clock [98,99]. Monitoring deviations between biological and chronological age helps to study development and aging across the lifespan [100]. Horvath [101], Hannum [93] and PhenoAge [102] epigenetic clocks serve as markers of ND [102–106], with the first of these showing the strongest correlation between epigenetics and chronological age [107].

Histone modifications can be used as potential aging MBM; however, the heterogeneity of the studies and the animal models limited the applicability of the findings. For highly abundant transcription activation mark H3K4me3 [48], the decrease in its level was shown to correlate with an extended lifespan in *Caenorhabditis elegans* [49], while an increase has been linked with AA and a reduced lifespan in *Drosophila melanogaster* [50]. For the heterochromatin-associated histone transcription repression mark H3K9me3 that is gradually lost during human and mouse aging in haematopoietic stem cells [51], the most significant changes occurred in the repressive regions in *C. elegans* [52] and models of AA [53]. The role of the repressive histone mark H3K27me3 associated with transcriptional silencing [54] in aging is controversial, as studies have shown both an increase and a decrease in its level during aging [55–59].

Increased H4K20me3 and H3K4me3 and decreased H3K9me1 and H3K27me3 histone modification levels have been described as common age-associated epigenetic marks [108–110]. In AD, an increase in the gene activation-related histone mark H3K4me3 was shown in both the CK-p25 tauopathy mouse model and in the hippocampus of AD patients [111,112]. Other histone methylation marks identified in AD patients' brain included: H4K20me2, H3K4me2, H3K27me3, H3K79me1, H3K79me2, H3K36me2, H4K20me3, H3K27me1 and

H3K56me1 [113,114]. In addition to methylation, histone acetylation marks H3K9ac, H3K14ac and H4K16ac have been shown to be associated with aging and AD [110,111,113–115]. Among other ND-associated histone modifications in AD, histone phosphorylation H4S47p and H3S10p [chaput2016potential] and histone ubiquitination H2BK120ub were reported [114,116,117]. The exact regulatory modes of both the modifications and mechanisms of their interaction and interplay with other factors in such complex processes as aging and ND are still unclear and require further systematic research.

Non-coding RNAs (ncRNAs) are used as aging MBM [118–121]. Long non-coding (lncRNA), for example, the growth-arrest-specific transcript 5 (*GAS5*), plays a significant role in cell proliferation and apoptosis [122–124], and its down-regulation leads to the phosphorylation of the tau protein in ND [125,126]. Long intergenic brain cytoplasmic RNA 1 (*BCYRN1*) expressed in the dendritic domains of neurons is down-regulated in aging [127].

MicroRNAs (miRNAs) mediate brain aging through the regulation of gene expression, impact neuronal plasticity and influence tau protein metabolism [128–137]. The regulation of *MiR145a* and *MiR-375* was shown to be age-dependent in mouse brains [138–140]. The *MIR29* family, *MIR339-5p*, *MIR195* and *MIR107*, regulate the expression of beta-secretase 1 that is responsible for proteolysis of the amyloid precursor protein [141–146]. Interestingly, *Mir34* was shown to play a protective role in *Drosophila* [147], and *MIR144/MIR451* was found to regulate ADAM metallopeptidase domain 10 in AD [148]. Over 20 miRNAs secreted into the cerebrospinal fluid by hypothalamic stem cells were found to control the aging rate in mice [149], which is especially important as the hypothalamus was placed at the base of human brain aging [150]. As most miRNA studies have been conducted on non-human models, their relevance to human data needs to be verified; hence, future studies will be required to define the roles of miRNAs [151].

Circular RNAs (circRNAs) are a recently described type of ncRNA with age-dependent expression in skeletal muscles [152] and an abundance in the brain [153], playing a role in ND through their interaction with miRNAs. For example, *CIRS-7* potentially functions as a sponge for *MIR7-1* [154] and its level is dramatically reduced in the AD brain [155]. Cerebral circRNAs are associated with neurotransmitter function, synaptic activities and neuronal maturation and target the expression and availability of specific age-related mRNAs in the brain. At least four circRNAs were found to be involved in postoperative neurocognitive disorders [156]. Another study revealed nearly 1200 cerebral circRNAs in a rat aging model [157]; however, these circRNAs still await their complete characterisation.

3. Aging of Organs and Systems beyond Neurodegeneration

Aging affects organs and systems with different rates of change; therefore, the AA concept needs to be adjusted when applied to individual organs. For example, ovarian aging implies a loss of follicle numbers and decreased oocyte viability. Typically, an accelerated decline in fertility begins around the age of 38 years and continues until the climacteric [158]; however, a non-uniform decrease in follicle numbers results in a large variation in menopause onset. The BA of the male reproduction system can also be assessed by fertility, but the arrest of reproductive capacity is reversible in older men, with lifestyle and disease factors prevailing over other determinants of aging [159]. In mice, oxidative stress, inflammation, DNA damage and de novo mutations accelerate testicular aging [160–163], while enhancing antioxidant enzyme activities with growth differentiation factor 11 protects the testes [164]. A progressive age-related drop in Leydig and Sertoli cell function [165], testicular size [166] or testosterone levels was demonstrated in older elite men [167]; however, no decrease in testicle size or the levels of testosterone was observed in a cohort of older elite men without comorbidities [168,169].

The cumulative effect of disease rather than age may account for changes in the male reproductive system throughout life. Obviously, chronological age does not always determine reproductive BA, and adopting the AA concept to the reproductive system is not allowed, which illustrates the challenges in assessing BA at the organ and system levels.

Sex hormones that affect fertility are part of the endocrine system. Data on the susceptibility of the endocrine system and metabolism to aging differ between organs and sex. The hypothalamic–pituitary–testicular axis in men does not undergo dramatic chronobiological changes, accounting for only 35–50% of men over 80 with reduced testosterone levels [170,171]. Conversely, diabetes mellitus and obesity predispose to accelerated adipose tissue dysfunction, affecting telomere length [172,173]. Adrenal and thyroid functions undergo less prominent age-related changes than their hypothalamic regulation [174]; therefore, assessing BA from hormonal findings is challenging. With aging, hormone activity decreases and endocrine alterations are established [175]. BA is affected by the level of glycosylated haemoglobin, glucose, triglycerides, and low-density and total cholesterol [176–178]. The modulation of these parameters, lifestyle and environmental factors can prevent or contribute to AA [179]. The effectiveness of hormone replacement therapy for aging reversal is questionable though [180].

Environmental and endocrinological factors affect the BA of connective tissue. The status of the skeletal system reflects the individual endocrine profile and micronutrient balance [181–183] as well as environmental and occupational attainments [184]. For example, bone resorption in astronauts prevails over its formation due to the effects of microgravity; however, bone density normalises after the flight [185,186]. Skin elasticity serves as a marker of aging, whose rate can be modified due to estrogen deficiency, metabolic alterations and exogenous factors (burns) [187–191]. Fibroblasts constitute a natural cell stock that allows skin rejuvenation, repair and decelerated aging [192]. In connective tissue, a combinatory effect of internal and external factors determines BA more accurately than the chronological one [193,194]. Therefore, the inability to account for the decreased aging rate reveals the weakness of the AA concept.

Studies in other systems have also reported the reversibility of age-related changes in them. For example, physical training can rejuvenate the respiratory system by expanding the alveolar space. However, studies on these issues did not comprehensively evaluate the BA of the lung since the impact of muscle atrophy on the results in the spirometry test was not considered [195–197]. Lifestyle changes (e.g., calorie restriction and physical activity) could also reverse aging in patients with early stages of chronic kidney disease [198]. Another example is shown by the discovered potential to rejuvenate the kidneys with up-regulation of the *Klotho* gene [199]. These evidences speak for a limited generalisation of the AA concept. AA affects various systems and cross-organ communication. The interaction between systems can impede the atrophy of an organ through compensatory mechanisms in other organs. Several studies have demonstrated the role of the central nervous system in reversing the aging of other systems and organs [200–202]. Endocrine and cardiovascular diseases promote renal aging [203]. Conversely, kidney transplantation can revive other parts of the body [204,205]. The characterisation of organ- and system-specific aging processes is challenging and will require combinatory approaches that are largely missing in the AA concept.

4. Limitations of the AA Concept

The AA concept is closely linked with and needs to be considered within the context of individual capacities and personalised structure–functional reserve mechanisms (Figure 2). The generalized term “structure-functional reserves” is introduced to approach the observed variability in multiple structure–functional parameters at different levels (expression of genetically and epigenetically regulated genes, number, viability and functionality of cells, number of synapses and intercellular contacts, secretion of cytokines, potency of physiological responses, etc.) in a population or group of subjects. The description of this term could be further developed and linked to the statistical distributions of measurable biological parameters in the population and to the norm of reaction resulting from the influence of environmental factors on trait variation.

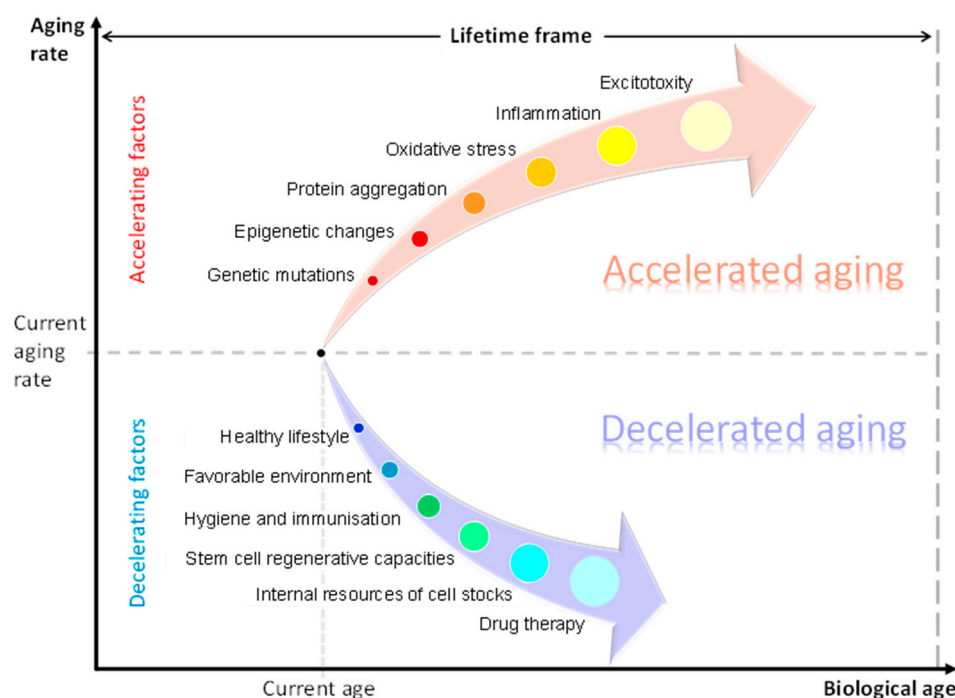


Figure 2. Factors influencing the rate of aging. Factors accelerating aging include: genetic mutations, epigenetic changes, protein aggregation, oxidative stress, inflammation and excitotoxicity. Factors decelerating aging include: a healthy lifestyle, favourable environment, hygiene and immunisation, stem cell regenerative capacities, internal resources of cell stocks and drug therapy.

Physiological reserves reflect the remaining capacity of an organ to perform its function. Aging and diseases lead to atrophy, reducing the number of cells and supracellular structures [206,207]. In the context of brain aging, its physiological cognitive reserve is assumed by the level of education, occupational and environmental attainments and the performance of cognitive tests [206]. Reversible forms of mild cognitive impairment (MCI) and dementia represent clinical examples of restoring individual reserve potential that are not in line with AA theory [208,209]. Neural compensation in the elderly leads to the formation of secondary brain networks [210], which decelerate the aging of the brain [206,211]. Reversion to MCI in elderly patients has been reported to be as a result of specific lifestyle activities and cognitive stimulation throughout life [212,213].

Appraising age requires an accurate estimation of individual reserves that account for biological and chronological age differences. In neuroscience, machine learning models establish an association between the number of years lived in good health and brain-imaging data with an accuracy of 2.1–4.9 years [214,215]. An individual's brain age can also be calculated as the difference between chronological age and the predicted BA [216]. In obstetrics, the evaluation of gynaecological status takes into account both reproductive health and potential fertility. Therefore, an overall BA depends on the reserve capacities of individual systems and organs [217,218].

Variance in individual reserve capacities complicates the precise assessment of BA. Brain AA criteria are unclear since normal aging indicators are still missing [219], and no reference curves for brain changes have been created yet. Moreover, the rate of aging is hard to assess due to the lack of a standardised methodology that could take into account individual reserve potentials. Furthermore, methodological discrepancies lead to contradictory findings between different studies that vary between laboratories. For example, published studies report that AD results in the addition of 1.5 years to the brain age; MCI adds 1 year; multiple sclerosis (MS), 0.41 years; Parkinson's disease (PD), 3 years; and schizophrenia, 5.5 years. However, the last two pathologies impact cognition in a milder and slower way than AD [220–222]. Other studies have reported an added brain

age of between 6 and 9 years in AD, which is more consistent with the findings on MCI and PD [223].

The weaknesses and limitations of certain studies show the need for caution when assessing results. For example, the studies on age-related brain atrophy commonly have a cross-sectional design that is less accurate compared to the longitudinal one [224]. Many studies are based on small non-representative cohorts [225–227]; therefore, the usage of the resulting mathematical models is low. Another challenge for the AA concept is due to the focus on middle-aged adults and elderly patients in certain brain aging studies. These studies should not ignore individual prenatal pathologies and childhood trauma affecting the brain's health and BA [228]. Applying the concept of AA to localised degeneration is complicated since different parts of the brain age unevenly [229]. For example, in localised ND, the BA assessment reflects the level of damage to the most vulnerable brain parts (e.g., substantia nigra and ruber nuclei in PD) [230–232]; however, one should also consider the brain resources that can minimise the atrophy effects [233]. In systematic ND, the brain ages faster than within localised ND [234,235], showing an apparent difference in the speed of atrophic changes [236].

Contemporary neuroscience lacks a clear explanation of the interaction between different causal factors due to the polyetiological nature of ND. It is still unclear whether chronic diseases lead to or result from ND [237,238] since the genetic, environmental and lifestyle factors interact in an undefined way [18,239,240]. Several articles have revealed a misalignment between dementia risk, cognitive performance and MBM levels [241,242]. Another disadvantage of AA studies is the inability to account for the influence of medications used by participants on study results [243]. Last but not least, AA represents a diagnostic but not pathognomonic signature in ND and in psychiatric diseases: schizophrenia, bipolar disorder and major depressive disorder [222,244,245], since the entire range of symptoms observed in these patients cannot be explained by brain aging only [222,246,247].

Finally, drug therapy could extend reserve abilities, for instance, in psychiatric conditions related to MCI and brain AA. Antidepressant medication has been reported to help convert MCI to true reversible MCI [248]. Certain cognitive disorders have demonstrated a reversible pattern in cognitive performance upon treatment [248,249]. Sex hormone replacement in ND reduced the risk, delayed the onset and slowed the progression of cognitive impairment [250]. Antioxidant-based therapy also alleviated the severity of the disease [251,252]. Recent ND studies have described a number of novel therapeutic options including specific antibodies, inducers of cell proliferation and NAD⁺ supplementation that are able to target mitophagy, protein aggregation and cellular senescence [9,18,253]. The observed treatment effects question the irreversible changes claimed by the AA concept.

5. Recommendations for Further Development and Improvement of AA Concept

5.1. Statistical Models

Building highly accurate machine learning models is the most common solution to distinguishing AA from normal aging. Recent articles suggest a set of approaches for improving the quality of these statistical solutions by reviewing the research data on MBM, the identification of potential gaps or inconsistencies, the incorporation of additional data sources or adjusting of research methodology, and the conduction of more rigorous statistical analyses, which will reveal any trends or correlations in the data that may have been overlooked. These suggestions have been commonly made by peers and experts who can provide valuable insights and suggestions. The following is the list of parameters that should be controlled to improve the diagnostic model of accelerated aging based on the use of specific molecular biomarkers:

- *Sample size.* The number of individuals in the study can affect the statistical power of the analysis, and larger sample sizes generally provide more robust results.
- *Biomarker types.* The choice of biomarkers can impact the diagnostic model used, as different types of biomarkers may require different statistical analyses.

- *Age range.* The age range of the study population can influence the types of biomarkers identified, as some biomarkers may be more prevalent in certain age groups.
- *Data normalization.* Normalization of the data is critical to ensure that data collection or processing differences do not affect the analysis.
- *Statistical methods.* The choice of statistical methods used can impact the sensitivity and specificity of the analysis, and different methods may be more appropriate for different types of data.

Careful consideration of these parameters will be critical in developing diagnostic models based on specific molecular biomarkers of accelerated aging. Recommendations on future developments of diagnostic models also include the functional characterisation of organ-tissue-specific, single-cell-specific and disease-specific molecular clocks, the integration of epigenetics into diverse large longitudinal studies, the exploration of additional epigenomic marks of aging, and the establishing and generation of data in robust non-human aging models.

5.2. Molecular Clocks

Consideration of organ-specific and tissue-specific aging molecular clocks in non-human models would further resolve the complexity of aging processes. Several useful molecular clocks in mice have already been reported [254], including those specific to organs and tissues: the liver [255–257], lung [255,256], blood [256,258], heart and cortex [255], adipose, kidney, muscle [256], and multi-tissue [259].

5.3. Single-Cell Epigenomics

The study of aging at single-cell resolution represents an important direction in aging science. In particular, the variation in gene expression and the corresponding changes in aging tissues and organs [260,261] suggest that single-cell methods will be needed for a detailed analysis of accelerated aging. For instance, a rise in cell-to-cell variability with age (“epigenomic noise”) associated with methylation increase in both H3K4me3 and H3K27me3 and with transcriptional heterogeneity was shown in immune cells in blood [262] and in muscle stem cells [263], respectively. Although arranging epigenetic clocks at the single-cell level will be technically challenging, new emerging methods [264,265] and deep-learning-based computer algorithms [266–268] may help to construct them.

5.4. New Epigenetic Biomarkers

The search for new epigenetic marks of aging represents another challenge and opens up new exciting research opportunities. The connections between aging and DNA modifications other than methylation are puzzling. Nevertheless, the signs of such associations are obvious. For example, deregulation of histone H4 acetylation (H4K12) [269] and accumulation of histone variant H2A.Z [270] with age were observed in a mouse hippocampus. Changes related to the aging-related acetylation of H3 (H3K9ac) and H4 (H4K16ac) histones were found in the brain of AD patients [113,271]. Furthermore, investigations on the modulation of histone acetylation by SIRT6 HDAC linked to longevity in mammals could lead to potential pharmacological developments to target AA [272–274].

5.5. Consideration of Ageotypes

Diagnostic models can be adjusted to different personalised physiological subsets of aging, ageotypes [44]. This approach considers various factors such as genetics, molecular clock parameters, other epigenetic changes, lifestyle habits and environmental exposures that may influence individual aging rates. Researchers can further adjust the diagnostic model of accelerated aging by incorporating numerous potential biomarkers of aging and health metrics [44,275,276] to more accurately classify ageotypes and monitor the effectiveness of future interventions on each subset.

5.6. Genetic Predisposition to AA

Certain diseases can be considered as clinically important models of human genetic predisposition to AA. In particular, the phenotypes of genome instability disorders that result from autosomal recessive mutations are associated with ineffective genome maintenance systems, deficiencies in DNA helicase activity or an aberrant nuclear architecture. Three groups of these disorders include the sunlight hypersensitivity disorders: *Xeroderma pigmentosum* (XP), Cockayne syndrome (CS) and trichothiodystrophy; the ionizing radiation hypersensitivity disorders including Ataxia telangiectasia (AT) and Nijmegen breakage syndrome (NBS); and the progeroid disorders including Werner syndrome, Hutchinson–Gilford progeria syndrome, Bloom syndrome, Rothmund–Thompson syndrome and Fanconi anemia [277–279].

5.7. Application of AA Animal Models

The emergence of new animal models exhibiting aging-related features (accelerated senescence, damage of nuclear envelope, increased accumulation of genomic lesions) can significantly contribute to AA research [280]. Well-developed mouse aging models provide the possibility of testing interventions and modulators [254]. For example, acceleration of the epigenetic clock by a high-fat diet, and the effects of caloric restriction and rapamycin were demonstrated in mice models [255,257]. Emerging new animal aging models include a vertebrate with the shortest captive lifespan, killifish (*Nothobranchius furzeri*) [281–287], longevity models such as naked mole rats (*Heterocephalus glaber*, *Fukomys mechowii*) [288–290], Brandt’s bat (*Myotis brandtii*) [291–293], olm (*Proteus anguinus*) [294–296], bivalve (*Arctica islandica*) [297,298] and non-aging organisms: Hydra (*Hydra vulgaris*/*Hydra magnipapillata*) [299–302] and Planaria (*Schmidtea mediterranea*) [303–305]. The use of these models can provide the generation of new robust aging data.

6. Conclusions

- i. The concept of an increased rate of age-related changes has certain weaknesses and limitations that are considered in the current review. In particular, so far, no unified methodology and terminology has been established in the field. The studies that justify the AA concept have too low sample sizes. Some age-related changes appear to be reversible under certain conditions.
- ii. Aging MBM help to estimate the aging rate increase due to a developed pathology or the exhaustion of individual reserves. A large variety of MBM candidates in different combinations can be associated with the aging brain; however, their validation, clinical interpretation and use in disease subtyping remain a challenge.
- iii. Activation of the regenerative mechanisms, and restoring metabolic and energy molecular reserves with novel therapeutic options could be potential ways to decelerate aging in the CNS. For example, sex hormone replacement, antioxidant-based and target therapy, and environmental and lifestyle factors’ improvement may delay ND. Future longitudinal studies could provide clinics and society with more options to prevent AA and slow the aging rate.

7. Afterword: Aging Science History and Theories

Several theories have been postulated to explain the possible biological meaning or evolutionary role of aging [306]: the evolutionary advantage of the species (1890s, Weisman); the accumulated mutation theory (1952, Medawar); the antagonistic pleiotropy (1957, Williams); the replicative senescence (1965, Hayflick); and the disposable soma theory (1972, Kirkwood). Furthermore, the causative theories of aging can be arranged in two groups: (I) Genetic (programmed) and (II) Stochastic (damage) theories. Programmed theories include programmed longevity theory, endocrine theory and immunological theory. Stochastic theories include wear-and-tear theory, rate of living theory, cross-linking theory, free radicals theory and somatic DNA damage theory [307].

Finally, the theories of aging can be classified by biological level and divided into: molecular level theories including gene regulation, codon restriction, error catastrophe, somatic mutation and dysdifferentiation theories; cellular level theories including cellular senescence–telomere theory, free radical theory, wear-and-tear theory and apoptosis theory; and system level theories including neuroendocrine theory, immunologic theory and rate of living theory [308].

The first publications of AA experiments performed at the Laboratories of the Rockefeller Institute for Medical Research date back to the 1920s with “normal aging” and “rate of aging” terms applied to the effects of light on *Drosophila* inbred in the dark [309,310]. Since then, the numbers of references on “aging”, “aging rate” and “AA” has reached 614,132; 56,088 and 21,401, respectively [311].

In 1928, the Professor of Neurology of the Columbia University, Frederick Tilney, published the work “The aging of the human brain”, where, in particular, the AD patient brain was compared to the normally aged one in the diagnostic context of the number of plaques. In this work, he also stated the abundance of senile plaques in all human brains after the age of 90 years, the influence of unfavourable factors and diseases on the brain and the importance of aging brain research. Recorded a century ago, his words are worth repeating today: “It is amazing how little general or particular interest man has shown in the most important organ of his body and life. Up to the present time he has devoted relatively little attention and much less capital to the understanding of that part of his machinery which is the secret of his success and the only hope for his future progress, if not his actual salvation. . . The ridiculous stupidity of annually consecrating appalling sums of money to the savage purposes of destruction should in time shock human intelligence out of patronizing such futilities and into wiser realizations. Certainly, one liberally supported and effective brain institute would prove an incomparably more profitable investment for civilization than the most powerful battle fleet that ever sailed the seas.” (Tilney, 1928 [312]).

Author Contributions: Y.S., S.A.A., B.S.E. and M.L. contributed to the conceptual idea of the paper. Y.S., K.N.-V.G. and R.H. formulated the objectives. Y.S., N.V.K., D.M. and K.L. wrote the manuscript. N.V.K., D.M. and K.L. prepared the figures for data presentation. N.V.K., G.L.S., D.S., A.K., S.M. and F.I. contributed to the literature review. All authors contributed to the writing of the original draft article. N.V.K. and B.S.E. performed the review and editing. All authors have read and agreed to the published version of the manuscript.

Funding: The study was supported by the ASPIRE, the technology program management pillar of Abu Dhabi’s Advanced Technology Research Council (ATRC), via the ASPIRE Precision Medicine Research Institute Abu Dhabi (VRI-20-10).

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

The following abbreviations are used in this manuscript:

5-mC	5-methylcytosine
AA	accelerated aging
AAG	gene related to accelerated aging
AD	Alzheimer’s disease
BA	biological age
BM	biomarker
circRNA	circular RNA
HC	healthy control
lncRNA	long non-coding RNA
MBM	molecular biomarkers
MCI	mild cognitive impairment
miRNA	microRNA
ncRNA	non-coding RNA
ND	neurodegeneration

NDG	gene that predispose to neurodegeneration
NV	neurovascular
NVU	neurovascular unit
PD	Parkinson's disease

References

- Isaev, N.K.; Stelmashook, E.V.; Genrikhs, E.E. Neurogenesis and brain aging. *Rev. Neurosci.* **2019**, *30*, 573–580. [[CrossRef](#)] [[PubMed](#)]
- Brivio, P.; Paladini, M.S.; Racagni, G.; Riva, M.A.; Calabrese, F.; Molteni, R. From healthy aging to frailty: In search of the underlying mechanisms. *Curr. Med. Chem.* **2019**, *26*, 3685–3701. [[CrossRef](#)] [[PubMed](#)]
- Feltes, B.C.; de Faria Poloni, J.; Bonatto, D. Development and aging: Two opposite but complementary phenomena. *Aging Health-A Syst. Biol. Perspect.* **2015**, *40*, 74–84.
- Bogeska, R.; Mikecin, A.M.; Kaschutnig, P.; Fawaz, M.; Büchler-Schäff, M.; Le, D.; Ganuza, M.; Vollmer, A.; Paffenholz, S.V.; Asada, N.; et al. Inflammatory exposure drives long-lived impairment of hematopoietic stem cell self-renewal activity and accelerated aging. *Cell Stem Cell* **2022**, *29*, 1273–1284. [[CrossRef](#)] [[PubMed](#)]
- Adelman, E.R.; Figueroa, M.E. Human hematopoiesis: Aging and leukemogenic risk. *Curr. Opin. Hematol.* **2021**, *28*, 57. [[CrossRef](#)]
- Hooten, N.N.; Pacheco, N.L.; Smith, J.T.; Evans, M.K. The accelerated aging phenotype: The role of race and social determinants of health on aging. *Ageing Res. Rev.* **2022**, *73*, 101536. [[CrossRef](#)]
- Forrester, S.N.; Zmora, R.; Schreiner, P.J.; Jacobs, D.R., Jr.; Roger, V.L.; Thorpe, R.J., Jr.; Kiefe, C.I. Accelerated aging: A marker for social factors resulting in cardiovascular events? *SSM-Popul. Health* **2021**, *13*, 100733. [[CrossRef](#)]
- Hamczyk, M.R.; Nevado, R.M.; Baretino, A.; Fuster, V.; Andres, V. Biological versus chronological aging: Jacc focus seminar. *J. Am. Coll. Cardiol.* **2020**, *75*, 919–930. [[CrossRef](#)]
- Vaquer-Alicea, J.; Diamond, M.I. Propagation of protein aggregation in neurodegenerative diseases. *Annu. Rev. Biochem.* **2019**, *88*, 785–810. [[CrossRef](#)]
- Armada-Moreira, A.; Gomes, J.I.; Pina, C.C.; Savchak, O.K.; Gonçalves-Ribeiro, J.; Rei, N.; Pinto, S.; Morais, T.P.; Martins, R.S.; Ribeiro, F.F.; et al. Going the extra (synaptic) mile: Excitotoxicity as the road toward neurodegenerative diseases. *Front. Cell. Neurosci.* **2020**, *14*, 90. [[CrossRef](#)]
- Mehta, A.; Prabhakar, M.; Kumar, P.; Deshmukh, R.; Sharma, P. Excitotoxicity: Bridge to various triggers in neurodegenerative disorders. *Eur. J. Pharmacol.* **2013**, *698*, 6–18. [[CrossRef](#)] [[PubMed](#)]
- Margolick, J.B.; Ferrucci, L. Accelerating aging research: How can we measure the rate of biologic aging? *Exp. Gerontol.* **2015**, *64*, 78–80. [[CrossRef](#)] [[PubMed](#)]
- Melzer, D.; Pilling, L.C.; Ferrucci, L. The genetics of human ageing. *Nat. Rev. Genet.* **2020**, *21*, 88–101. [[CrossRef](#)] [[PubMed](#)]
- Miller, M.W.; Sadeh, N. Traumatic stress, oxidative stress and post-traumatic stress disorder: Neurodegeneration and the accelerated-aging hypothesis. *Mol. Psychiatry* **2014**, *19*, 1156–1162. [[CrossRef](#)] [[PubMed](#)]
- Ghosh, C.; De, A. Basics of aging theories and disease related aging-an overview. *PharmaTutor* **2017**, *5*, 16–23.
- Wadhwa, R.; Gupta, R.; Maurya, P.K. Oxidative stress and accelerated aging in neurodegenerative and neuropsychiatric disorder. *Curr. Pharm. Des.* **2018**, *24*, 4711–4725. [[CrossRef](#)]
- Bersani, F.S.; Mellon, S.H.; Reus, V.I.; Wolkowitz, O.M. Accelerated aging in serious mental disorders. *Curr. Opin. Psychiatry* **2019**, *32*, 381. [[CrossRef](#)]
- Hou, Y.; Dan, X.; Babbar, M.; Wei, Y.; Hasselbalch, S.G.; Croteau, D.L.; Bohr, V.A. Ageing as a risk factor for neurodegenerative disease. *Nat. Rev. Neurol.* **2019**, *15*, 565–581. [[CrossRef](#)]
- Wang, X.; Ma, Z.; Cheng, J.; Lv, Z. A genetic program theory of aging using an RNA population model. *Ageing Res. Rev.* **2014**, *13*, 46–54. [[CrossRef](#)]
- Kovacs, G.G. Concepts and classification of neurodegenerative diseases. *Handb. Clin. Neurol.* **2018**, *145*, 301–307.
- Sanz, A.; Stefanatos, R.K. The mitochondrial free radical theory of aging: A critical view. *Curr. Aging Sci.* **2008**, *1*, 10–21. [[CrossRef](#)] [[PubMed](#)]
- Libertini, G.; Shubernetskaya, O.; Corbi, G.; Ferrara, N. Is evidence supporting the subtelomere–telomere theory of aging? *Biochemistry* **2021**, *86*, 1526–1539. [[CrossRef](#)]
- Xie, L.; Wu, S.; He, R.; Li, S.; Lai, X.; Wang, Z. Identification of epigenetic dysregulation gene markers and immune landscape in kidney renal clear cell carcinoma by comprehensive genomic analysis. *Front. Immunol.* **2022**, *13*, 901662. [[CrossRef](#)] [[PubMed](#)]
- Růžička, M.; Kulhánek, P.; Radová, L.; Čechová, A.; Špačková, N.; Fajkusová, L.; Rěblová, K. Dna mutation motifs in the genes associated with inherited diseases. *PLoS ONE* **2017**, *12*, e0182377. [[CrossRef](#)] [[PubMed](#)]
- Korb, M.K.; Kimonis, V.E.; Mozaffar, T. Multisystem proteinopathy: Where myopathy and motor neuron disease converge. *Muscle Nerve* **2021**, *63*, 442–454. [[CrossRef](#)]
- Barja, G. The mitochondrial free radical theory of aging. *Prog. Mol. Biol. Transl. Sci.* **2014**, *127*, 1–27.
- Amorim, J.A.; Coppotelli, G.; Rolo, A.P.; Palmeira, C.M.; Ross, J.M.; Sinclair, D.A. Mitochondrial and metabolic dysfunction in aging and age-related diseases. *Nat. Rev. Endocrinol.* **2022**, *18*, 243–258. [[CrossRef](#)]

28. Esmaeili, Y.; Yarjanli, Z.; Pakniya, F.; Bidram, E.; Los, M.J.; Eshraghi, M.; Klionsky, D.J.; Ghavami, S.; Zarrabi, A. Targeting autophagy, oxidative stress, and ER stress for neurodegenerative diseases treatment. *J. Control. Release* **2022**, *345*, 147–175. [[CrossRef](#)]
29. Pomatto, L.C.; Davies, K.J. Adaptive homeostasis and the free radical theory of ageing. *Free Radic. Biol. Med.* **2018**, *124*, 420–430. [[CrossRef](#)]
30. Simpson, D.J.; Chandra, T. Epigenetic age prediction. *Aging Cell* **2021**, *20*, e13452. [[CrossRef](#)]
31. Schaeffer, S.; Iadecola, C. Revisiting the neurovascular unit. *Nat. Neurosci.* **2021**, *24*, 1198–1209. [[CrossRef](#)] [[PubMed](#)]
32. Campisi, J. Cancer, aging and cellular senescence. *In Vivo* **2000**, *14*, 183–188. [[PubMed](#)]
33. Zlokovic, B.V. New therapeutic targets in the neurovascular pathway in Alzheimer's disease. *Neurotherapeutics* **2008**, *5*, 409–414. [[CrossRef](#)]
34. Hu, X.; De Silva, T.M.; Chen, J.; Faraci, F.M. Cerebral vascular disease and neurovascular injury in ischemic stroke. *Circ. Res.* **2017**, *120*, 449–471. [[CrossRef](#)] [[PubMed](#)]
35. Lähtenvuo, J.; Rosenzweig, A. Effects of aging on angiogenesis. *Circ. Res.* **2012**, *110*, 1252–1264. [[CrossRef](#)]
36. Montagne, A.; Barnes, S.R.; Sweeney, M.D.; Halliday, M.R.; Sagare, A.P.; Zhao, Z.; Toga, A.W.; Jacobs, R.E.; Liu, C.Y.; Amezcua, L.; et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* **2015**, *85*, 296–302. [[CrossRef](#)]
37. Nelson, A.R.; Sweeney, M.D.; Sagare, A.P.; Zlokovic, B.V. Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta Mol. Basis Dis.* **2016**, *1862*, 887–900. [[CrossRef](#)]
38. Wilhelm, I.; Nyúl-Tóth, Á.; Kozma, M.; Farkas, A.E.; Krizbai, I.A. Role of pattern recognition receptors of the neurovascular unit in inflamm-aging. *Am. J. Physiol. Heart Circ. Physiol.* **2017**, *313*, H1000–H1012. [[CrossRef](#)]
39. Zhou, Z.D.; Wang, D.Q.; Tan, E.K. The role of neurovascular unit in neurodegeneration. *Front. Cell. Neurosci.* **2022**, *16*, 870631. [[CrossRef](#)]
40. Spitzer, D.; Guérit, S.; Puetz, T.; Khel, M.I.; Armbrust, M.; Dunst, M.; Macas, J.; Zinke, J.; Devraj, G.; Jia, X.; et al. Profiling the neurovascular unit unveils detrimental effects of osteopontin on the blood–brain barrier in acute ischemic stroke. *Acta Neuropathol.* **2022**, *144*, 305–337. [[CrossRef](#)]
41. Jeong, H.W.; Diéguez-Hurtado, R.; Arf, H.; Song, J.; Park, H.; Kruse, K.; Sorokin, L.; Adams, R.H. Single-cell transcriptomics reveals functionally specialized vascular endothelium in brain. *eLife* **2022**, *11*, e57520. [[CrossRef](#)] [[PubMed](#)]
42. Zhao, Y.; Xie, Y.Z.; Liu, Y.S. Accelerated aging-related transcriptome alterations in neurovascular unit cells in the brain of Alzheimer's disease. *Front. Aging Neurosci.* **2022**, *14*, 949074. [[CrossRef](#)]
43. Xia, X.; Chen, W.; McDermott, J.; Han, J.D.J. Molecular and phenotypic biomarkers of aging. *F1000Research* **2017**, *6*, 860. [[CrossRef](#)] [[PubMed](#)]
44. Ahadi, S.; Zhou, W.; Schüssler-Fiorenza Rose, S.M.; Sailani, M.R.; Contrepolis, K.; Avina, M.; Ashland, M.; Brunet, A.; Snyder, M. Personal aging markers and ageotypes revealed by deep longitudinal profiling. *Nat. Med.* **2020**, *26*, 83–90. [[CrossRef](#)] [[PubMed](#)]
45. Song, Z.; von Figura, G.; Liu, Y.; Kraus, J.M.; Torrice, C.; Dillon, P.; Rudolph-Watabe, M.; Ju, Z.; Kestler, H.A.; Sanoff, H.; et al. Lifestyle impacts on the aging-associated expression of biomarkers of DNA damage and telomere dysfunction in human blood. *Aging Cell* **2010**, *9*, 607–615. [[CrossRef](#)]
46. Horvath, S.; Zhang, Y.; Langfelder, P.; Kahn, R.S.; Boks, M.P.; van Eijk, K.; van den Berg, L.H.; Ophoff, R.A. Aging effects on DNA methylation modules in human brain and blood tissue. *Genome Biol.* **2012**, *13*, 1–18. [[CrossRef](#)]
47. Day, K.; Waite, L.L.; Thalacker-Mercer, A.; West, A.; Bamman, M.M.; Brooks, J.D.; Myers, R.M.; Absher, D. Differential DNA methylation with age displays both common and dynamic features across human tissues that are influenced by CpG landscape. *Genome Biol.* **2013**, *14*, 1–19. [[CrossRef](#)]
48. Greer, E.L.; Shi, Y. Histone methylation: A dynamic mark in health, disease and inheritance. *Nat. Rev. Genet.* **2012**, *13*, 343–357. [[CrossRef](#)]
49. Greer, E.L.; Maures, T.J.; Hauswirth, A.G.; Green, E.M.; Leeman, D.S.; Maro, G.S.; Han, S.; Banko, M.R.; Gozani, O.; Brunet, A. Members of the h3k4 trimethylation complex regulate lifespan in a germline-dependent manner in *C. elegans*. *Nature* **2010**, *466*, 383–387. [[CrossRef](#)]
50. Li, L.; Greer, C.; Eisenman, R.N.; Secombe, J. Essential functions of the histone demethylase lid. *PLoS Genet.* **2010**, *6*, e1001221. [[CrossRef](#)]
51. Djegloul, D.; Kuranda, K.; Kuzniak, I.; Barbieri, D.; Naguibneva, I.; Choisy, C.; Bories, J.C.; Dosquet, C.; Pla, M.; Vanneaux, V.; et al. Age-associated decrease of the histone methyltransferase SUV39H1 in hSC perturbs heterochromatin and B lymphoid differentiation. *Stem Cell Rep.* **2016**, *6*, 970–984. [[CrossRef](#)] [[PubMed](#)]
52. Li, C.L.; Pu, M.; Wang, W.; Chaturvedi, A.; Emerson, F.J.; Lee, S.S. Region-specific h3k9me3 gain in aged somatic tissues in *Caenorhabditis elegans*. *PLoS Genet.* **2021**, *17*, e1009432. [[CrossRef](#)] [[PubMed](#)]
53. Lee, J.H.; Kim, E.W.; Croteau, D.L.; Bohr, V.A. Heterochromatin: An epigenetic point of view in aging. *Exp. Mol. Med.* **2020**, *52*, 1466–1474. [[CrossRef](#)] [[PubMed](#)]
54. Cao, R.; Wang, L.; Wang, H.; Xia, L.; Erdjument-Bromage, H.; Tempst, P.; Jones, R.S.; Zhang, Y. Role of histone H3 lysine 27 methylation in Polycomb-group silencing. *Science* **2002**, *298*, 1039–1043. [[CrossRef](#)] [[PubMed](#)]
55. Siebold, A.P.; Banerjee, R.; Tie, F.; Kiss, D.L.; Moskowicz, J.; Harte, P.J. Polycomb repressive complex 2 and trithorax modulate *Drosophila* longevity and stress resistance. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 169–174. [[CrossRef](#)]

56. Ni, Z.; Ebata, A.; Alipanahramandi, E.; Lee, S.S. Two set domain containing genes link epigenetic changes and aging in *Caenorhabditis elegans*. *Aging Cell* **2012**, *11*, 315–325. [[CrossRef](#)]
57. Maures, T.J.; Greer, E.L.; Hauswirth, A.G.; Brunet, A. The h3k27 demethylase *utx-1* regulates *C. elegans* lifespan in a germline-independent, insulin-dependent manner. *Aging Cell* **2011**, *10*, 980–990. [[CrossRef](#)]
58. Liu, L.; Cheung, T.H.; Charville, G.W.; Hurgo, B.M.; Leavitt, T.; Shih, J.; Brunet, A.; Rando, T.A. Chromatin modifications as determinants of muscle stem cell quiescence and chronological aging. *Cell Rep.* **2013**, *4*, 189–204. [[CrossRef](#)]
59. Baumgart, M.; Groth, M.; Priebe, S.; Savino, A.; Testa, G.; Dix, A.; Ripa, R.; Spallotta, F.; Gaetano, C.; Ori, M.; et al. RNA-seq of the aging brain in the short-lived fish *N. furzeri*—conserved pathways and novel genes associated with neurogenesis. *Aging Cell* **2014**, *13*, 965–974. [[CrossRef](#)]
60. Peters, M.J.; Joehanes, R.; Pilling, L.C.; Schurmann, C.; Conneely, K.N.; Powell, J.; Reinmaa, E.; Sutphin, G.L.; Zhernakova, A.; Schramm, K.; et al. The transcriptional landscape of age in human peripheral blood. *Nat. Commun.* **2015**, *6*, 8570. [[CrossRef](#)]
61. Li, X.; Khanna, A.; Li, N.; Wang, E. Circulatory mir-34a as an RNA-based, noninvasive biomarker for brain aging. *Aging* **2011**, *3*, 985. [[CrossRef](#)] [[PubMed](#)]
62. Dhahbi, J.M. Circulating small noncoding RNAs as biomarkers of aging. *Ageing Res. Rev.* **2014**, *17*, 86–98. [[CrossRef](#)] [[PubMed](#)]
63. Grammatikakis, I.; Panda, A.C.; Abdelmohsen, K.; Gorospe, M. Long noncoding RNAs (lncRNAs) and the molecular hallmarks of aging. *Aging* **2014**, *6*, 992. [[CrossRef](#)] [[PubMed](#)]
64. Kour, S.; Rath, P.C. Long noncoding RNAs in aging and age-related diseases. *Ageing Res. Rev.* **2016**, *26*, 1–21. [[CrossRef](#)]
65. Finkel, D.; Pedersen, N.L.; Plomin, R.; McClearn, G.E. Longitudinal and cross-sectional twin data on cognitive abilities in adulthood: The Swedish Adoption/Twin Study of Aging. *Dev. Psychol.* **1998**, *34*, 1400. [[CrossRef](#)]
66. Reynolds, C.A.; Finkel, D. A meta-analysis of heritability of cognitive aging: Minding the “missing heritability” gap. *Neuropsychol. Rev.* **2015**, *25*, 97–112. [[CrossRef](#)]
67. Blauwendraat, C.; Pletnikova, O.; Geiger, J.T.; Murphy, N.A.; Abramzon, Y.; Rudow, G.; Mamais, A.; Sabir, M.S.; Crain, B.; Ahmed, S.; et al. Genetic analysis of neurodegenerative diseases in a pathology cohort. *Neurobiol. Aging* **2019**, *76*, 214.e1–214.e9. [[CrossRef](#)]
68. Cochran, J.N.; Geier, E.G.; Bonham, L.W.; Newberry, J.S.; Amaral, M.D.; Thompson, M.L.; Lasseigne, B.N.; Karydas, A.M.; Roberson, E.D.; Cooper, G.M.; et al. Non-coding and loss-of-function coding variants in *tet2* are associated with multiple neurodegenerative diseases. *Am. J. Hum. Genet.* **2020**, *106*, 632–645. [[CrossRef](#)]
69. Cirulli, E.T.; Lasseigne, B.N.; Petrovski, S.; Sapp, P.C.; Dion, P.A.; Leblond, C.S.; Couthouis, J.; Lu, Y.F.; Wang, Q.; Krueger, B.J.; et al. Exome sequencing in amyotrophic lateral sclerosis identifies risk genes and pathways. *Science* **2015**, *347*, 1436–1441. [[CrossRef](#)]
70. Chung, S.J.; Kim, M.J.; Kim, J.; Kim, Y.J.; You, S.; Koh, J.; Kim, S.Y.; Lee, J.H. Exome array study did not identify novel variants in Alzheimer’s disease. *Neurobiol. Aging* **2014**, *35*, 1958.e13–1958.e14. [[CrossRef](#)]
71. Nikolic Perkovic, M.; Pivac, N. Genetic Markers of Alzheimer’s Disease. *Adv. Exp. Med. Biol.* **2019**, *1192*, 27–52. [[PubMed](#)]
72. Song, W.; Hooli, B.; Mullin, K.; Jin, S.C.; Cella, M.; Ulland, T.K.; Wang, Y.; Tanzi, R.E.; Colonna, M. Alzheimer’s disease-associated *trem2* variants exhibit either decreased or increased ligand-dependent activation. *Alzheimer’s Dement.* **2017**, *13*, 381–387. [[CrossRef](#)] [[PubMed](#)]
73. Ruiz, A.; Dols-Icardo, O.; Bullido, M.J.; Pastor, P.; Rodríguez-Rodríguez, E.; López de Munain, A.; de Pancorbo, M.M.; Pérez-Tur, J.; Alvarez, V.; Antonell, A.; et al. Assessing the role of the *trem2* p. r47h variant as a risk factor for Alzheimer’s disease and frontotemporal dementia. *Neurobiol. Aging* **2014**, *35*, 444.e1–444.e4. [[CrossRef](#)] [[PubMed](#)]
74. Mehrjoo, Z.; Najmabadi, A.; Abedini, S.S.; Mohseni, M.; Kamali, K.; Najmabadi, H.; Khorram Khorshid, H.R. Association study of the *trem2* gene and identification of a novel variant in exon 2 in Iranian patients with late-onset Alzheimer’s disease. *Med. Princ. Pract.* **2015**, *24*, 351–354. [[CrossRef](#)] [[PubMed](#)]
75. Guerreiro, R.; Wojtas, A.; Bras, J.; Carrasquillo, M.; Rogaeve, E.; Majounie, E.; Cruchaga, C.; Sassi, C.; Kauwe, J.S.; Younkin, S.; et al. *Trem2* variants in Alzheimer’s disease. *N. Engl. J. Med.* **2013**, *368*, 117–127. [[CrossRef](#)]
76. Jonsson, T.; Stefansson, H.; Steinberg, S.; Jonsson, P.V.; Snaedal, J.; Bjornsson, S.; Huttenlocher, J.; Levey, A.I.; Lah, J.J.; et al. Variant of *trem2* associated with the risk of Alzheimer’s disease. *N. Engl. J. Med.* **2013**, *368*, 107–116. [[CrossRef](#)]
77. Jiang, T.; Tan, L.; Chen, Q.; Tan, M.S.; Zhou, J.S.; Zhu, X.C.; Lu, H.; Wang, H.F.; Zhang, Y.D.; Yu, J.T. A rare coding variant in *trem2* increases risk for Alzheimer’s disease in Han Chinese. *Neurobiol. Aging* **2016**, *42*, 217.e1–217.e3. [[CrossRef](#)]
78. Jin, S.C.; Carrasquillo, M.M.; Benitez, B.A.; Skorupa, T.; Carrell, D.; Patel, D.; Lincoln, S.; Krishnan, S.; Kachadourian, M.; Reitz, C.; et al. *Trem2* is associated with increased risk for Alzheimer’s disease in African Americans. *Mol. Neurodegener.* **2015**, *10*, 1–7. [[CrossRef](#)]
79. Berge, G.; Sando, S.B.; Rongve, A.; Aarsland, D.; White, L.R. Onset of dementia with Lewy bodies is delayed for carriers of the apolipoprotein E $\epsilon 2$ genotype in a Norwegian cohort. *Mov. Disord.* **2014**, *29*, S220.
80. Calvo, A.; Chiò, A. Sclerosi laterale amiotrofica come modello di gestione interdisciplinare. *SALUTE E SOCIETÀ* **2015**, *3*, 173–184. [[CrossRef](#)]
81. Borroni, B.; Ferrari, F.; Galimberti, D.; Nacmias, B.; Barone, C.; Bagnoli, S.; Fenoglio, C.; Piaceri, I.; Archetti, S.; Bonvicini, C.; et al. Heterozygous *trem2* mutations in frontotemporal dementia. *Neurobiol. Aging* **2014**, *35*, 934.e7–934.e10. [[CrossRef](#)] [[PubMed](#)]
82. Rayaprolu, S.; Mullen, B.; Baker, M.; Lynch, T.; Finger, E.; Seeley, W.W.; Hatanpaa, K.J.; Lomen-Hoerth, C.; Kertesz, A.; Bigio, E.H.; et al. *Trem2* in neurodegeneration: Evidence for association of the p. r47h variant with frontotemporal dementia and Parkinson’s disease. *Mol. Neurodegener.* **2013**, *8*, 1–5. [[CrossRef](#)] [[PubMed](#)]

83. Cady, J.; Koval, E.D.; Benitez, B.A.; Zaidman, C.; Jockel-Balsarotti, J.; Allred, P.; Baloh, R.H.; Ravits, J.; Simpson, E.; Appel, S.H.; et al. Trem2 variant p. r47h as a risk factor for sporadic amyotrophic lateral sclerosis. *JAMA Neurol.* **2014**, *71*, 449–453. [[CrossRef](#)] [[PubMed](#)]
84. Slattery, C.F.; Beck, J.A.; Harper, L.; Adamson, G.; Abdi, Z.; Uphill, J.; Campbell, T.; Drueyeh, R.; Mahoney, C.J.; Rohrer, J.D.; et al. R47h trem2 variant increases risk of typical early-onset Alzheimer’s disease but not of prion or frontotemporal dementia. *Alzheimer’s Dement.* **2014**, *10*, 602–608. [[CrossRef](#)] [[PubMed](#)]
85. Gonzalez Murcia, J.D.; Schmutz, C.; Munger, C.; Perkes, A.; Gustin, A.; Peterson, M.; Ebbert, M.T.; Norton, M.C.; Tschanz, J.T.; Munger, R.G.; et al. Assessment of trem2 rs75932628 association with Alzheimer’s disease in a population-based sample: The cache county study. *Neurobiol. Aging* **2013**, *34*, 2889. [[CrossRef](#)]
86. Walton, R.L.; Soto-Ortolaza, A.I.; Murray, M.E.; Lorenzo-Betancor, O.; Ogaki, K.; Heckman, M.G.; Rayaprolu, S.; Rademakers, R.; Ertekin-Taner, N.; Uitti, R.J.; et al. Trem2 p. r47h substitution is not associated with dementia with lewy bodies. *Neurol. Genet.* **2016**, *2*, e85. [[CrossRef](#)]
87. Sun, J.; Zhu, Z.; Chen, K.; Wei, D.; Li, X.; Li, H.; Zhang, J.; Chen, X.; Chen, Y.; Zhang, Z. Apoe ϵ 4 allele accelerates age-related multi-cognitive decline and white matter damage in non-demented elderly. *Aging* **2020**, *12*, 12019. [[CrossRef](#)]
88. Goel, N.; Karir, P.; Garg, V.K. Role of DNA methylation in human age prediction. *Mech. Ageing Dev.* **2017**, *166*, 33–41. [[CrossRef](#)]
89. Jung, M.; Pfeifer, G.P. Aging and DNA methylation. *BMC Biol.* **2015**, *13*, 1–8. [[CrossRef](#)]
90. Lister, R.; Pelizzola, M.; Downen, R.H.; Hawkins, R.D.; Hon, G.; Tonti-Filippini, J.; Nery, J.R.; Lee, L.; Ye, Z.; Ngo, Q.M.; et al. Human DNA methylomes at base resolution show widespread epigenomic differences. *Nature* **2009**, *462*, 315–322. [[CrossRef](#)]
91. Jones, P.A. Functions of DNA methylation: Islands, start sites, gene bodies and beyond. *Nat. Rev. Genet.* **2012**, *13*, 484–492. [[CrossRef](#)] [[PubMed](#)]
92. Zampieri, M.; Ciccarone, F.; Calabrese, R.; Franceschi, C.; Bürkle, A.; Caiafa, P. Reconfiguration of DNA methylation in aging. *Mech. Ageing Dev.* **2015**, *151*, 60–70. [[CrossRef](#)] [[PubMed](#)]
93. Hannum, G.; Guinney, J.; Zhao, L.; Zhang, L.; Hughes, G.; Sada, S.; Klotzle, B.; Bibikova, M.; Fan, J.B.; Gao, Y.; et al. Genome-wide methylation profiles reveal quantitative views of human aging rates. *Mol. Cell* **2013**, *49*, 359–367. [[CrossRef](#)]
94. Fraga, M.F.; Ballestar, E.; Paz, M.F.; Ropero, S.; Setien, F.; Ballestar, M.L.; Heine-Suñer, D.; Cigudosa, J.C.; Urioste, M.; Benitez, J.; et al. Epigenetic differences arise during the lifetime of monozygotic twins. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 10604–10609. [[CrossRef](#)] [[PubMed](#)]
95. Ryan, C.P. “Epigenetic clocks”: Theory and applications in human biology. *Am. J. Hum. Biol.* **2021**, *33*, e23488. [[CrossRef](#)]
96. Martino, D.; Loke, Y.J.; Gordon, L.; Ollikainen, M.; Cruickshank, M.N.; Saffery, R.; Craig, J.M. Longitudinal, genome-scale analysis of dna methylation in twins from birth to 18 months of age reveals rapid epigenetic change in early life and pair-specific effects of discordance. *Genome Biol.* **2013**, *14*, R42. [[CrossRef](#)]
97. Bjornsson, H.T.; Sigurdsson, M.I.; Fallin, M.D.; Irizarry, R.A.; Aspelund, T.; Cui, H.; Yu, W.; Rongione, M.A.; Ekström, T.J.; Harris, T.B.; et al. Intra-individual change over time in dna methylation with familial clustering. *JAMA* **2008**, *299*, 2877–2883. [[CrossRef](#)]
98. Wynford-Thomas, D. Telomeres, p53 and cellular senescence. *Oncol Res.* **1996**, *8*, 387–398.
99. von Zglinicki, T. Telomeres: Influencing the rate of aging. *Ann. N. Y. Acad. Sci.* **1998**, *854*, 318–327. [[CrossRef](#)]
100. Teschendorff, A.E.; West, J.; Beck, S. Age-associated epigenetic drift: Implications, and a case of epigenetic thrift? *Hum. Mol. Genet.* **2013**, *22*, R7–R15. [[CrossRef](#)]
101. Horvath, S. Dna methylation age of human tissues and cell types. *Genome Biol.* **2013**, *14*, R115. [[CrossRef](#)] [[PubMed](#)]
102. Levine, M.E.; Lu, A.T.; Quach, A.; Chen, B.H.; Assimes, T.L.; Bandinelli, S.; Hou, L.; Baccarelli, A.A.; Stewart, J.D.; Li, Y.; et al. An epigenetic biomarker of aging for lifespan and healthspan. *Aging* **2018**, *10*, 573–591. [[CrossRef](#)] [[PubMed](#)]
103. Marioni, R.E.; Shah, S.; McRae, A.F.; Ritchie, S.J.; Muniz-Terrera, G.; Harris, S.E.; Gibson, J.; Redmond, P.; Cox, S.R.; Pattie, A.; et al. The epigenetic clock is correlated with physical and cognitive fitness in the lothian birth cohort 1936. *Int. J. Epidemiol.* **2015**, *44*, 1388–1396. [[CrossRef](#)] [[PubMed](#)]
104. Levine, M.E.; Lu, A.T.; Bennett, D.A.; Horvath, S. Epigenetic age of the pre-frontal cortex is associated with neuritic plaques, amyloid load, and Alzheimer’s disease related cognitive functioning. *Aging* **2015**, *7*, 1198. [[CrossRef](#)]
105. Horvath, S.; Langfelder, P.; Kwak, S.; Aaronson, J.; Rosinski, J.; Vogt, T.F.; Eszes, M.; Faull, R.L.; Curtis, M.A.; Waldvogel, H.J.; et al. Huntington’s disease accelerates epigenetic aging of human brain and disrupts dna methylation levels. *Aging* **2016**, *8*, 1485. [[CrossRef](#)]
106. Grodstein, F.; Lemos, B.; Yu, L.; Klein, H.U.; Iatrou, A.; Buchman, A.S.; Shireby, G.L.; Mill, J.; Schneider, J.A.; De Jager, P.L.; et al. The association of epigenetic clocks in brain tissue with brain pathologies and common aging phenotypes. *Neurobiol. Dis.* **2021**, *157*, 105428. [[CrossRef](#)]
107. Grodstein, F.; Lemos, B.; Yu, L.; Iatrou, A.; De Jager, P.L.; Bennett, D.A. Characteristics of epigenetic clocks across blood and brain tissue in older women and men. *Front. Neurosci.* **2021**, *14*, 555307. [[CrossRef](#)]
108. Fraga, M.F.; Esteller, M. Epigenetics and aging: The targets and the marks. *Trends Genet.* **2007**, *23*, 413–418. [[CrossRef](#)]
109. Han, S.; Brunet, A. Histone methylation makes its mark on longevity. *Trends Cell Biol.* **2012**, *22*, 42–49. [[CrossRef](#)]
110. López-Otín, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. *Cell* **2013**, *153*, 1194–1217. [[CrossRef](#)]
111. Gjonjeska, E.; Pfenning, A.R.; Mathys, H.; Quon, G.; Kundaje, A.; Tsai, L.H.; Kellis, M. Conserved epigenomic signals in mice and humans reveal immune basis of Alzheimer’s disease. *Nature* **2015**, *518*, 365–369. [[CrossRef](#)] [[PubMed](#)]

112. Cao, Q.; Wang, W.; Williams, J.B.; Yang, F.; Wang, Z.J.; Yan, Z. Targeting histone K4 trimethylation for treatment of cognitive and synaptic deficits in mouse models of Alzheimer's disease. *Sci. Adv.* **2020**, *6*, eabc8096. [[CrossRef](#)] [[PubMed](#)]
113. Nativio, R.; Donahue, G.; Berson, A.; Lan, Y.; Amlie-Wolf, A.; Tuzer, F.; Toledo, J.B.; Gosai, S.J.; Gregory, B.D.; Torres, C.; et al. Dysregulation of the epigenetic landscape of normal aging in Alzheimer's disease. *Nat. Neurosci.* **2018**, *21*, 497–505. [[CrossRef](#)]
114. Santana, D.A.; Smith MD, A.C.; Chen, E.S. Histone modifications in alzheimer's disease. *Genes* **2023**, *14*, 347. [[CrossRef](#)]
115. Tang, B.; Dean, B.; Thomas, E. Disease-and age-related changes in histone acetylation at gene promoters in psychiatric disorders. *Transl. Psychiatry* **2011**, *1*, e64. [[CrossRef](#)]
116. Chaput, D.; Kirouac, L.; Stevens, S.M., Jr.; Padmanabhan, J. Potential role of PCTAIRE-2, PCTAIRE-3 and P-Histone H4 in amyloid precursor protein-dependent Alzheimer pathology. *Oncotarget* **2016**, *7*, 8481. [[CrossRef](#)] [[PubMed](#)]
117. Ogawa, O.; Zhu, X.; Lee, H.G.; Raina, A.; Obrenovich, M.E.; Bowser, R.; Smith, M.A. Ectopic localization of phosphorylated histone H3 in Alzheimer's disease: A mitotic catastrophe? *Acta Neuropathol.* **2003**, *105*, 524–528. [[CrossRef](#)] [[PubMed](#)]
118. D'haene, E.; Vergult, S. Interpreting the impact of noncoding structural variation in neurodevelopmental disorders. *Genet. Med.* **2021**, *23*, 34–46. [[CrossRef](#)] [[PubMed](#)]
119. Sherazi, S.A.M.; Abbasi, A.; Jamil, A.; Uzair, M.; Ikram, A.; Qamar, S.; Olamide, A.A.; Arshad, M.; Fried, P.J.; Ljubicavljovic, M.; et al. Molecular hallmarks of long non-coding RNAs in aging and its significant effect on aging-associated diseases. *Neural Regen. Res.* **2023**, *18*, 959–968.
120. Wang, D.Q.; Fu, P.; Yao, C.; Zhu, L.S.; Hou, T.Y.; Chen, J.G.; Lu, Y.; Liu, D.; Zhu, L.Q. Long non-coding RNAs, novel culprits, or bodyguards in neurodegenerative diseases. *Mol. Ther. Nucleic Acids* **2018**, *10*, 269–276. [[CrossRef](#)]
121. Mishra, P.; Kumar, S. Association of lncRNA with regulatory molecular factors in brain and their role in the pathophysiology of schizophrenia. *Metab. Brain Dis.* **2021**, *36*, 849–858. [[CrossRef](#)] [[PubMed](#)]
122. Coccia, E.M.; Cicala, C.; Charlesworth, A.; Ciccarelli, C.; Rossi, G.B.; Philipson, L.; Sorrentino, V. Regulation and expression of a growth arrest-specific gene (gas5) during growth, differentiation, and development. *Mol. Cell. Biol.* **1992**, *12*, 3514–3521.
123. Pickard, M.; Mourtada-Maarabouni, M.; Williams, G. Long non-coding RNA gas5 regulates apoptosis in prostate cancer cell lines. *Biochim. Biophys. Acta Mol. Basis Dis.* **2013**, *1832*, 1613–1623. [[CrossRef](#)] [[PubMed](#)]
124. Mourtada-Maarabouni, M.; Pickard, M.; Hedge, V.; Farzaneh, F.; Williams, G. Gas5, a non-protein-coding RNA, controls apoptosis and is downregulated in breast cancer. *Oncogene* **2009**, *28*, 195–208. [[CrossRef](#)] [[PubMed](#)]
125. Tang, S.; Buchman, A.S.; De Jager, P.L.; Bennett, D.A.; Epstein, M.P.; Yang, J. Novel variance-component TWAS method for studying complex human diseases with applications to Alzheimer's dementia. *PLoS Genet.* **2021**, *17*, e1009482. [[CrossRef](#)]
126. Liang, W.S.; Dunckley, T.; Beach, T.G.; Grover, A.; Mastroeni, D.; Walker, D.G.; Caselli, R.J.; Kukull, W.A.; McKeel, D.; Morris, J.C.; et al. Gene expression profiles in anatomically and functionally distinct regions of the normal aged human brain. *Physiol. Genom.* **2007**, *28*, 311–322. [[CrossRef](#)]
127. Mus, E.; Hof, P.R.; Tiedge, H. Dendritic bc200 RNA in aging and in Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 10679–10684. [[CrossRef](#)]
128. Maoz, R.; Garfinkel, B.P.; Soreq, H. Alzheimer's disease and ncRNAs. *Neuroepigenomics Aging Dis.* **2017**, *978*, 337–361.
129. Fiore, R.; Khudayberdiev, S.; Saba, R.; Schrott, G. Micro-RNA function in the nervous system. *Prog. Mol. Biol. Transl. Sci.* **2011**, *102*, 47–100.
130. Goodall, E.F.; Heath, P.R.; Bandmann, O.; Kirby, J.; Shaw, P.J. Neuronal dark matter: The emerging role of microRNAs in neurodegeneration. *Front. Cell. Neurosci.* **2013**, *7*, 178. [[CrossRef](#)]
131. Dickson, J.R.; Kruse, C.; Montagna, D.R.; Finsen, B.; Wolfe, M.S. Alternative polyadenylation and mir-34 family members regulate tau expression. *J. Neurochem.* **2013**, *127*, 739–749. [[CrossRef](#)] [[PubMed](#)]
132. Smith, P.Y.; Hernandez-Rapp, J.; Jolivet, F.; Lecours, C.; Bisht, K.; Goupil, C.; Dorval, V.; Parsi, S.; Morin, F.; Planel, E.; et al. Mir-132/212 deficiency impairs tau metabolism and promotes pathological aggregation in vivo. *Hum. Mol. Genet.* **2015**, *24*, 6721–6735. [[CrossRef](#)] [[PubMed](#)]
133. Santa-Maria, I.; Alaniz, M.E.; Renwick, N.; Cela, C.; Fulga, T.A.; Van Vactor, D.; Tuschl, T.; Clark, L.N.; Shelanski, M.L.; McCabe, B.D.; et al. Dysregulation of microRNA-219 promotes neurodegeneration through post-transcriptional regulation of tau. *J. Clin. Investig.* **2015**, *125*, 681–686. [[CrossRef](#)] [[PubMed](#)]
134. Hébert, S.S.; Papadopoulou, A.S.; Smith, P.; Galas, M.C.; Planel, E.; Silahiroglu, A.N.; Sergeant, N.; Buée, L.; De Strooper, B. Genetic ablation of dicer in adult forebrain neurons results in abnormal tau hyperphosphorylation and neurodegeneration. *Hum. Mol. Genet.* **2010**, *19*, 3959–3969. [[CrossRef](#)] [[PubMed](#)]
135. Cai, Z.; Zhao, Y.; Zhao, B. Roles of glycogen synthase kinase 3 in Alzheimer's disease. *Curr. Alzheimer Res.* **2012**, *9*, 864–879. [[CrossRef](#)] [[PubMed](#)]
136. Mohamed, J.S.; Lopez, M.A.; Boriek, A.M. Mechanical stretch up-regulates microRNA-26a and induces human airway smooth muscle hypertrophy by suppressing glycogen synthase kinase-3 β . *J. Biol. Chem.* **2010**, *285*, 29336–29347. [[CrossRef](#)]
137. Zhao, Z.B.; Wu, L.; Xiong, R.; Wang, L.L.; Zhang, B.; Wang, C.; Li, H.; Liang, L.; Chen, S.D. MicroRNA-922 promotes tau phosphorylation by downregulating ubiquitin carboxy-terminal hydrolase 11 (uchl1) expression in the pathogenesis of Alzheimer's disease. *Neuroscience* **2014**, *275*, 232–237. [[CrossRef](#)]
138. Law, P.T.; Ching, A.K.; Chan, A.W.; Wong, Q.W.; Wong, C.K.; To, K.F.; Wong, N. Mir-145 modulates multiple components of the insulin-like growth factor pathway in hepatocellular carcinoma. *Carcinogenesis* **2012**, *33*, 1134–1141. [[CrossRef](#)]

139. El Ouaamari, A.; Baroukh, N.; Martens, G.A.; Lebrun, P.; Pipeleers, D.; Van Obberghen, E. mir-375 targets 3-phosphoinositide-dependent protein kinase-1 and regulates glucose-induced biological responses in pancreatic β -cells. *Diabetes* **2008**, *57*, 2708–2717. [[CrossRef](#)]
140. Inukai, S.; de Lencastre, A.; Turner, M.; Slack, F. Novel microRNAs differentially expressed during aging in the mouse brain. *PLoS ONE* **2012**, *7*, e40028. [[CrossRef](#)]
141. Yang, G.; Song, Y.; Zhou, X.; Deng, Y.; Liu, T.; Weng, G.; Yu, D.; Pan, S. MicroRNA-29c targets β -site amyloid precursor protein-cleaving enzyme 1 and has a neuroprotective role in vitro and in vivo. *Mol. Med. Rep.* **2015**, *12*, 3081–3088. [[CrossRef](#)] [[PubMed](#)]
142. Lei, X.; Lei, L.; Zhang, Z.; Zhang, Z.; Cheng, Y. Downregulated mir-29c correlates with increased bace1 expression in sporadic Alzheimer's disease. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 1565. [[PubMed](#)]
143. Zong, Y.; Wang, H.; Dong, W.; Quan, X.; Zhu, H.; Xu, Y.; Huang, L.; Ma, C.; Qin, C. mir-29c regulates bace1 protein expression. *Brain Res.* **2011**, *1395*, 108–115. [[CrossRef](#)] [[PubMed](#)]
144. Hébert, S.S.; Horré, K.; Nicolai, L.; Bergmans, B.; Papadopoulou, A.S.; Delacourte, A.; De Strooper, B. MicroRNA regulation of Alzheimer's amyloid precursor protein expression. *Neurobiol. Dis.* **2009**, *33*, 422–428. [[CrossRef](#)]
145. Zhu, H.C.; Wang, L.M.; Wang, M.; Song, B.; Tan, S.; Teng, J.F.; Duan, D.X. MicroRNA-195 downregulates Alzheimer's disease amyloid- β production by targeting bace1. *Brain Res. Bull.* **2012**, *88*, 596–601. [[CrossRef](#)]
146. Wang, W.X.; Rajeev, B.W.; Stromberg, A.J.; Ren, N.; Tang, G.; Huang, Q.; Rigoutsos, I.; Nelson, P.T. The expression of microRNA mir-107 decreases early in Alzheimer's disease and may accelerate disease progression through regulation of β -site amyloid precursor protein-cleaving enzyme 1. *J. Neurosci.* **2008**, *28*, 1213–1223. [[CrossRef](#)]
147. Kennerdell, J.R.; Liu, N.; Bonini, N.M. Mir-34 inhibits polycomb repressive complex 2 to modulate chaperone expression and promote healthy brain aging. *Nat. Commun.* **2018**, *9*, 4188. [[CrossRef](#)]
148. Cheng, C.; Li, W.; Zhang, Z.; Yoshimura, S.; Hao, Q.; Zhang, C.; Wang, Z. MicroRNA-144 is regulated by activator protein-1 (ap-1) and decreases expression of Alzheimer disease-related a disintegrin and metalloprotease 10 (adam10). *J. Biol. Chem.* **2013**, *288*, 13748–13761. [[CrossRef](#)]
149. Zhang, Y.; Kim, M.S.; Jia, B.; Yan, J.; Zuniga-Hertz, J.P.; Han, C.; Cai, D. Hypothalamic stem cells control ageing speed partly through exosomal miRNAs. *Nature* **2017**, *548*, 52–57. [[CrossRef](#)]
150. Zhang, G.; Li, J.; Purkayastha, S.; Tang, Y.; Zhang, H.; Yin, Y.; Li, B.; Liu, G.; Cai, D. Hypothalamic programming of systemic ageing involving ikk- β , nf- κ b and gnrh. *Nature* **2013**, *497*, 211–216. [[CrossRef](#)]
151. Mohammed, C.P.D.; Park, J.S.; Nam, H.G.; Kim, K. MicroRNAs in brain aging. *Mech. Ageing Dev.* **2017**, *168*, 3–9. [[CrossRef](#)]
152. Abdelmohsen, K.; Panda, A.C.; De, S.; Grammatikakis, I.; Kim, J.; Ding, J.; Noh, J.H.; Kim, K.M.; Mattison, J.A.; de Cabo, R.; et al. Circular RNAs in monkey muscle: Age-dependent changes. *Ageing* **2015**, *7*, 903. [[CrossRef](#)]
153. Rybak-Wolf, A.; Stottmeister, C.; Glazar, P.; Jens, M.; Pino, N.; Giusti, S.; Hanan, M.; Behm, M.; Bartok, O.; Ashwal-Fluss, R.; et al. Circular RNAs in the mammalian brain are highly abundant, conserved, and dynamically expressed. *Mol. Cell* **2015**, *58*, 870–885. [[CrossRef](#)]
154. Hansen, T.B.; Jensen, T.I.; Clausen, B.H.; Bramsen, J.B.; Finsen, B.; Damgaard, C.K.; Kjems, J. Natural RNA circles function as efficient microRNA sponges. *Nature* **2013**, *495*, 384–388. [[CrossRef](#)] [[PubMed](#)]
155. Lukiw, W.J. Circular RNA (circRNA) in Alzheimer's disease (ad). *Front. Genet.* **2013**, *4*, 307. [[CrossRef](#)]
156. Bao, N.; Liu, J.; Peng, Z.; Zhang, R.; Ni, R.; Li, R.; Wu, J.; Liu, Z.; Pan, B. Identification of circRNA-miRNA-mRNA networks to explore the molecular mechanism and immune regulation of postoperative neurocognitive disorder. *Ageing* **2022**, *14*, 8374. [[CrossRef](#)] [[PubMed](#)]
157. Mahmoudi, E.; Fitzsimmons, C.; Geaghan, M.P.; Shannon Weickert, C.; Atkins, J.R.; Wang, X.; Cairns, M.J. Circular RNA biogenesis is decreased in postmortem cortical gray matter in schizophrenia and may alter the bioavailability of associated miRNA. *Neuropsychopharmacology* **2019**, *44*, 1043–1054. [[CrossRef](#)] [[PubMed](#)]
158. Broekmans, F.; Soules, M.; Fauser, B. Ovarian aging: Mechanisms and clinical consequences. *Endocr. Rev.* **2009**, *30*, 465–493. [[CrossRef](#)]
159. Frungieri, M.B.; Calandra, R.S.; Bartke, A.; Matzkin, M.E. Male and female gonadal ageing: Its impact on health span and life span. *Mech. Ageing Dev.* **2021**, *197*, 111519. [[CrossRef](#)]
160. De la Rochebrochard, E.; Thonneau, P. Paternal age 40 years: An important risk factor for infertility. *Am. J. Obstet. Gynecol.* **2003**, *189*, 901–905. [[CrossRef](#)]
161. Brahem, S.; Mehdi, M.; Elghezal, H.; Saad, A. The effects of male aging on semen quality, sperm dna fragmentation and chromosomal abnormalities in an infertile population. *J. Assist. Reprod. Genet.* **2011**, *28*, 425–432. [[CrossRef](#)] [[PubMed](#)]
162. Paul, C.; Robaire, B. Ageing of the male germ line. *Nat. Rev. Urol.* **2013**, *10*, 227–234. [[CrossRef](#)] [[PubMed](#)]
163. Frungieri, M.B.; Calandra, R.S.; Bartke, A.; Matzkin, M.E. Ageing and inflammation in the male reproductive tract. *Andrologia* **2018**, *50*, e13034. [[CrossRef](#)] [[PubMed](#)]
164. Zhou, Y.; Ni, S.; Li, C.; Song, L.; Zhang, S. Gonadal rejuvenation of mice by growth differentiation factor 11. *J. Gerontol. Ser. A* **2022**, *77*, 892–901. [[CrossRef](#)]
165. Mularoni, V.; Esposito, V.; Di Persio, S.; Vicini, E.; Spadetta, G.; Berloco, P.; Fanelli, F.; Mezzullo, M.; Pagotto, U.; Pelusi, C.; et al. Age-related changes in human Leydig cell status. *Hum. Reprod.* **2020**, *35*, 2663–2676. [[CrossRef](#)]

166. Mahmoud, A.; Goemaere, S.; El-Garem, Y.; Van Pottelbergh, I.; Comhaire, F.; Kaufman, J. Testicular volume in relation to hormonal indices of gonadal function in community-dwelling elderly men. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 179–184. [[CrossRef](#)]
167. Golan, R.; Scovell, J.M.; Ramasamy, R. Age-related testosterone decline is due to waning of both testicular and hypothalamic-pituitary function. *Aging Male* **2015**, *18*, 201–204. [[CrossRef](#)]
168. Handelsman, D.J.; Staraj, S. Testicular size: The effects of aging, malnutrition, and illness. *J. Androl.* **1985**, *6*, 144–151. [[CrossRef](#)]
169. Iacqua, A.; Izzo, G.; Emerenziani, G.P.; Baldari, C.; Aversa, A. Lifestyle and fertility: The influence of stress and quality of life on male fertility. *Reprod. Biol. Endocrinol.* **2018**, *16*, 1–11. [[CrossRef](#)]
170. Kaufman, J. Ageing of the hypothalamo-pituitary-testicular axis in men. *Horm. Res. Paediatr.* **1995**, *43*, 25–28.
171. Harman, S.M.; Metter, E.J.; Tobin, J.D.; Pearson, J.; Blackman, M.R. Longitudinal effects of aging on serum total and free testosterone levels in healthy men. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 724–731. [[CrossRef](#)] [[PubMed](#)]
172. Spinelli, R.; Parrillo, L.; Longo, M.; Florese, P.; Desiderio, A.; Zatterale, F.; Miele, C.; Raciti, G.A.; Beguinot, F. Molecular basis of ageing in chronic metabolic diseases. *J. Endocrinol. Investig.* **2020**, *43*, 1373–1389. [[CrossRef](#)] [[PubMed](#)]
173. Emami, M.; Agbaedeng, T.A.; Thomas, G.; Middeldorp, M.E.; Thiyagarajah, A.; Wong, C.X.; Elliott, A.D.; Gallagher, C.; Hendriks, J.M.L.; Lau, D.H.; et al. Accelerated biological aging secondary to cardiometabolic risk factors is a predictor of cardiovascular mortality: A systematic review and meta-analysis. *Can. J. Cardiol.* **2022**, *38*, 365–375. [[CrossRef](#)]
174. Russell, S.J.; Kahn, C.R. Endocrine regulation of ageing. *Nat. Rev. Mol. Cell Biol.* **2007**, *8*, 681–691. [[CrossRef](#)] [[PubMed](#)]
175. Van den Beld, A.W.; Kaufman, J.M.; Zillikens, M.C.; Lamberts, S.W.; Egan, J.M.; van der Lely, A.J. The physiology of endocrine systems with ageing. *Lancet Diabetes Endocrinol.* **2018**, *6*, 647–658. [[CrossRef](#)] [[PubMed](#)]
176. Park, J.; Cho, B.; Kwon, H.; Lee, C. Developing a biological age assessment equation using principal component analysis and clinical biomarkers of aging in Korean men. *Arch. Gerontol. Geriatr.* **2009**, *49*, 7–12. [[CrossRef](#)]
177. Nakamura, E.; Moritani, T.; Kanetaka, A. Effects of habitual physical exercise on physiological age in men aged 20–85 years as estimated using principal component analysis. *Eur. J. Appl. Physiol. Occup. Physiol.* **1996**, *73*, 410–418. [[CrossRef](#)]
178. Nakamura, E.; Moritani, T.; Kanetaka, A. Biological age versus physical fitness age in women. *Eur. J. Appl. Physiol. Occup. Physiol.* **1990**, *61*, 202–208. [[CrossRef](#)]
179. Nunn, A.V.; Bell, J.D.; Guy, G.W. Lifestyle-induced metabolic inflexibility and accelerated ageing syndrome: Insulin resistance, friend or foe? *Nutr. Metab.* **2009**, *6*, 1–26. [[CrossRef](#)]
180. Chahal, H.; Drake, W. The endocrine system and ageing. *J. Pathol. A J. Pathol. Soc. Great Br. Irel.* **2007**, *211*, 173–180. [[CrossRef](#)]
181. Chandra, A.; Rajawat, J. Skeletal aging and osteoporosis: Mechanisms and therapeutics. *Int. J. Mol. Sci.* **2021**, *22*, 3553. [[CrossRef](#)] [[PubMed](#)]
182. Gaffney-Stomberg, E.; Hughes, J.M.; Guerriere, K.I.; Staab, J.S.; Cable, S.J.; Bouxsein, M.L.; McClung, J.P. Once daily calcium (1000 mg) and vitamin D (1000 IU) supplementation during military training prevents increases in biochemical markers of bone resorption but does not affect tibial microarchitecture in army recruits. *Bone* **2022**, *155*, 116269. [[CrossRef](#)] [[PubMed](#)]
183. Wang, S.; Luo, Z.; Luo, H.; Li, Z.; Yuan, Z.; Tang, J.; Lin, L.; Du, Z.; Zhou, J.R. Effects of a calcium/vitamin D/zinc combination on anti-osteoporosis in ovariectomized rats. *J. Trace Elem. Med. Biol.* **2023**, *77*, 127138. [[CrossRef](#)]
184. Sfeir, J.G.; Drake, M.T.; Khosla, S.; Farr, J.N. Skeletal aging. *Mayo Clin. Proc.* **2022**, *97*, 1194–1208. [[CrossRef](#)] [[PubMed](#)]
185. Carter, M.I.; Hinton, P.S. Physical activity and bone health. *MO Med.* **2014**, *111*, 59.
186. Orwoll, E.S.; Adler, R.A.; Amin, S.; Binkley, N.; Lewiecki, E.M.; Petak, S.M.; Shapses, S.A.; Sinaki, M.; Watts, N.B.; Sibonga, J.D. Skeletal health in long-duration astronauts: Nature, assessment, and management recommendations from the NASA bone summit. *J. Bone Miner. Res.* **2013**, *28*, 1243–1255. [[CrossRef](#)]
187. Boismal, F.; Serron, K.; Dobos, G.; Zuelgaray, E.; Bensussan, A.; Michel, L. Skin aging: Pathophysiology and innovative therapies. *Med. Sci. M/S* **2020**, *36*, 1163–1172. [[CrossRef](#)]
188. Baumann, L. Skin ageing and its treatment. *J. Pathol. A J. Pathol. Soc. Great Br. Irel.* **2007**, *211*, 241–251. [[CrossRef](#)]
189. Kohl, E.; Steinbauer, J.; Landthaler, M.; Szeimies, R.M. Skin ageing. *J. Eur. Acad. Dermatol. Venereol.* **2011**, *25*, 873–884. [[CrossRef](#)]
190. Brincat, M.; Muscat Baron, Y.; Galea, R. Estrogens and the skin. *Climacteric* **2005**, *8*, 110–123. [[CrossRef](#)]
191. Park, H.Y.; Kim, J.H.; Jung, M.; Chung, C.H.; Hasham, R.; Park, C.S.; Choi, E.H. A long-standing hyperglycaemic condition impairs skin barrier by accelerating skin ageing process. *Exp. Dermatol.* **2011**, *20*, 969–974. [[CrossRef](#)] [[PubMed](#)]
192. Bonté, F.; Girard, D.; Archambault, J.C.; Desmoulière, A. Skin changes during ageing. *Biochem. Cell Biol. Ageing Pt II Clin. Sci.* **2019**, *91*, 249–280.
193. Shpichka, A.; Butnaru, D.; Bezrukov, E.A.; Sukhanov, R.B.; Atala, A.; Burdukovskii, V.; Zhang, Y.; Timashev, P. Skin tissue regeneration for burn injury. *Stem Cell Res. Ther.* **2019**, *10*, 1–16. [[CrossRef](#)] [[PubMed](#)]
194. Bae, C.Y.; Kang, Y.G.; Piao, M.H.; Cho, B.; Cho, K.H.; Park, Y.K.; Yu, B.Y.; Lee, S.W.; Kim, M.J.; Lee, S.H.; et al. Models for estimating the biological age of five organs using clinical biomarkers that are commonly measured in clinical practice settings. *Maturitas* **2013**, *75*, 253–260. [[CrossRef](#)] [[PubMed](#)]
195. Naskalova, S.; Shatilo, V.; Pizaruk, A.; Antoniuk-Shcheglova, I.; Bondarenko, O.; Bodretska, L.; Shapovalenko, I. Estimating the functional age of the respiratory system. *Ageing Longev.* **2022**, *3*, 71–76. [[CrossRef](#)]
196. Sprung, J.; Gajic, O.; Warner, D.O. Age related alterations in respiratory function—anesthetic considerations. *Can. J. Anesth.* **2006**, *53*, 1244. [[CrossRef](#)]
197. Charansonney, O.L. Physical activity and aging: A life-long story. *Discov. Med.* **2011**, *12*, 177–185.

198. Negri, A.L. The klotho gene: A gene predominantly expressed in the kidney is a fundamental regulator of aging and calcium/phosphorus metabolism. *J. Nephrol.* **2005**, *18*, 654–658.
199. Wei, S.Y.; Pan, S.Y.; Li, B.; Chen, Y.M.; Lin, S.L. Rejuvenation: Turning back the clock of aging kidney. *J. Formos. Med. Assoc.* **2020**, *119*, 898–906. [[CrossRef](#)]
200. Alcedo, J.; Flatt, T.; Pasyukova, E.G. The role of the nervous system in aging and longevity. *Front. Genet.* **2013**, *4*, 124. [[CrossRef](#)]
201. Bouchard, J.; Villeda, S.A. Aging and brain rejuvenation as systemic events. *J. Neurochem.* **2015**, *132*, 5–19. [[CrossRef](#)] [[PubMed](#)]
202. Kimura, K.; Ieda, M.; Kanazawa, H.; Yagi, T.; Tsunoda, M.; Ninomiya, S.; Kurosawa, H.; Yoshimi, K.; Mochizuki, H.; Yamazaki, K.; et al. Cardiac sympathetic rejuvenation: A link between nerve function and cardiac hypertrophy. *Circ. Res.* **2007**, *100*, 1755–1764. [[CrossRef](#)] [[PubMed](#)]
203. Martin, J.; Sheaff, M. Renal ageing. *J. Pathol. A J. Pathol. Soc. Great Br. Irel.* **2007**, *211*, 198–205. [[CrossRef](#)] [[PubMed](#)]
204. Harciarek, M.; Biedunkiewicz, B.; Lichodziejewska-Niemierko, M.; Dębska-Ślizień, A.; Rutkowski, B. Continuous cognitive improvement 1 year following successful kidney transplant. *Kidney Int.* **2011**, *79*, 1353–1360. [[CrossRef](#)] [[PubMed](#)]
205. Griva, K.; Thompson, D.; Jayasena, D.; Davenport, A.; Harrison, M.; Newman, S.P. Cognitive functioning pre-to post-kidney transplantation—A prospective study. *Nephrol. Dial. Transplant.* **2006**, *21*, 3275–3282. [[CrossRef](#)] [[PubMed](#)]
206. Stern, Y. Cognitive reserve in ageing and Alzheimer’s disease. *Lancet Neurol.* **2012**, *11*, 1006–1012. [[CrossRef](#)]
207. Amanvermez, R.; Tosun, M. An update on ovarian aging and ovarian reserve tests. *Int. J. Fertil. Steril.* **2016**, *9*, 411.
208. Chilovi, B.V.; Caratozzolo, S.; Mombelli, G.; Zanetti, M.; Rozzini, L.; Padovani, A. Does reversible mci exist? *Alzheimer’s Dement.* **2011**, *7*, 5. [[CrossRef](#)]
209. Vermunt, L.; van Paasen, A.J.L.; Teunissen, C.E.; Scheltens, P.; Visser, P.J.; Tijms, B.M.; Alzheimer’s Disease Neuroimaging Initiative. Alzheimer disease biomarkers may aid in the prognosis of mci cases initially reverted to normal. *Neurology* **2019**, *92*, e2699–e2705. [[CrossRef](#)]
210. Zarahn, E.; Rakitin, B.; Abela, D.; Flynn, J.; Stern, Y. Age-related changes in brain activation during a delayed item recognition task. *Neurobiol. Aging* **2007**, *28*, 784–798. [[CrossRef](#)]
211. Oosterhuis, E.J.; Slade, K.; May, P.J.; Nuttall, H.E. Towards an understanding of healthy cognitive ageing: The importance of lifestyle in cognitive reserve and the scaffolding theory of aging and cognition. *J. Gerontol. Ser. B* **2022**, *78*, 777–788. [[CrossRef](#)] [[PubMed](#)]
212. Shimada, H.; Doi, T.; Lee, S.; Makizako, H. Reversible predictors of reversion from mild cognitive impairment to normal cognition: A 4-year longitudinal study. *Alzheimer’s Res. Ther.* **2019**, *11*, 1–9. [[CrossRef](#)] [[PubMed](#)]
213. Valenzuela, M.J.; Sachdev, P.; Wen, W.; Chen, X.; Brodaty, H. Lifespan mental activity predicts diminished rate of hippocampal atrophy. *PLoS ONE* **2008**, *3*, e2598. [[CrossRef](#)] [[PubMed](#)]
214. Aycheh, H.M.; Seong, J.K.; Shin, J.H.; Na, D.L.; Kang, B.; Seo, S.W.; Sohn, K.A. Biological brain age prediction using cortical thickness data: A large scale cohort study. *Front. Aging Neurosci.* **2018**, *10*, 252. [[CrossRef](#)] [[PubMed](#)]
215. Franke, K.; Gaser, C. Ten years of brainage as a neuroimaging biomarker of brain aging: What insights have we gained? *Front. Neurol.* **2019**, *10*, 789. [[CrossRef](#)]
216. Elliott, M.L.; Belsky, D.W.; Knodt, A.R.; Ireland, D.; Melzer, T.R.; Poulton, R.; Ramrakha, S.; Caspi, A.; Moffitt, T.E.; Hariri, A.R. Brain-age in midlife is associated with accelerated biological aging and cognitive decline in a longitudinal birth cohort. *Mol. Psychiatry* **2021**, *26*, 3829–3838. [[CrossRef](#)]
217. Saxon, S.V.; Etten, M.J.; Perkins, E.A. *Physical Change and Aging: A Guide for Helping Professions*; Springer Publishing Company: Berlin/Heidelberg, Germany, 2021.
218. Berezina, T.N.; Rybtsov, S.A. Use of personal resources may influence the rate of biological aging depending on individual typology. *Eur. J. Investig. Health Psychol. Educ.* **2022**, *12*, 1793–1811. [[CrossRef](#)]
219. Bethlehem, R.A.I.; Seidlitz, J.; White, S.R.; Vogel, J.W.; Anderson, K.M.; Adamson, C.; Adler, S.; Alexopoulos, G.S.; Anagnostou, E.; Areces-Gonzalez, A. Brain charts for the human lifespan. *Nature* **2022**, *604*, 525–533. [[CrossRef](#)]
220. Gaser, C.; Franke, K.; Klöppel, S.; Koutsouleris, N.; Sauer, H.; Initiative, A.D.N. Brainage in mild cognitive impaired patients: Predicting the conversion to Alzheimer’s disease. *PLoS ONE* **2013**, *8*, e67346. [[CrossRef](#)]
221. Eickhoff, C.R.; Hoffstaedter, F.; Caspers, J.; Reetz, K.; Mathys, C.; Dogan, I.; Amunts, K.; Schnitzler, A.; Eickhoff, S.B. Advanced brain ageing in Parkinson’s disease is related to disease duration and individual impairment. *Brain Commun.* **2021**, *3*, fcab191. [[CrossRef](#)]
222. Koutsouleris, N.; Davatzikos, C.; Borgwardt, S.; Gaser, C.; Bottlender, R.; Frodl, T.; Falkai, P.; Riecher-Rössler, A.; Möller, H.J.; Reiser, M.; et al. Accelerated brain aging in schizophrenia and beyond: A neuroanatomical marker of psychiatric disorders. *Schizophr. Bull.* **2014**, *40*, 1140–1153. [[CrossRef](#)] [[PubMed](#)]
223. Franke, K.; Gaser, C. Longitudinal changes in individual brainage in healthy aging, mild cognitive impairment, and Alzheimer’s disease. *GeroPsych* **2012**, *25*, 235–245. [[CrossRef](#)]
224. Schill, R.I.; Frost, C.; Jenkins, R.; Whitwell, J.L.; Rossor, M.N.; Fox, N.C. A longitudinal study of brain volume changes in normal aging using serial registered magnetic resonance imaging. *Arch. Neurol.* **2003**, *60*, 989–994. [[CrossRef](#)]
225. Coffey, C.E.; Wilkinson, W.E.; Parashos, I.A.; Soady, S.A.; Sullivan, R.J.; Patterson, L.J.; Figiel, G.S.; Webb, M.C.; Spritzer, C.E.; Djang, W.T. Quantitative cerebral anatomy of the aging human brain: A cross-sectional study using magnetic resonance imaging. *Neurology* **1992**, *42*, 527. [[CrossRef](#)]

226. Good, C.D.; Johnsrude, I.S.; Ashburner, J.; Henson, R.N.; Friston, K.J.; Frackowiak, R.S. A voxel-based morphometric study of ageing in 465 normal adult human brains. *Neuroimage* **2001**, *14*, 21–36. [[CrossRef](#)] [[PubMed](#)]
227. Iwasaki, A.; Foxman, E.F.; Molony, R.D. Early local immune defences in the respiratory tract. *Nat. Rev. Immunol.* **2017**, *17*, 7–20. [[CrossRef](#)]
228. Sarkar, T.; Patro, N.; Patro, I.K. Cumulative multiple early life hits—a potent threat leading to neurological disorders. *Brain Res. Bull.* **2019**, *147*, 58–68. [[CrossRef](#)]
229. Hawkes, C.H.; Del Tredici, K.; Braak, H. A timeline for parkinson’s disease. *Park. Relat. Disord.* **2010**, *16*, 79–84. [[CrossRef](#)]
230. Turknnett, J.; Wood, T.R. Demand coupling drives neurodegeneration: A model of age-related cognitive decline and dementia. *Cells* **2022**, *11*, 2789. [[CrossRef](#)]
231. Schneider, C.B.; Donix, M.; Linse, K.; Werner, A.; Fauser, M.; Klingelhoefer, L.; Löhle, M.; von Kummer, R.; Reichmann, H.; Storch, A.; et al. Accelerated age-dependent hippocampal volume loss in parkinson disease with mild cognitive impairment. *Am. J. Alzheimer’s Dis. Other Dement.* **2017**, *32*, 313–319. [[CrossRef](#)]
232. Smith, S.M.; Elliott, L.T.; Alfaro-Almagro, F.; McCarthy, P.; Nichols, T.E.; Douaud, G.; Miller, K.L. Brain aging comprises many modes of structural and functional change with distinct genetic and biophysical associations. *eLife* **2020**, *9*, e52677. [[CrossRef](#)] [[PubMed](#)]
233. Bayati, A.; Berman, T. Localized vs. systematic neurodegeneration: A paradigm shift in understanding neurodegenerative diseases. *Front. Syst. Neurosci.* **2017**, *11*, 62. [[CrossRef](#)] [[PubMed](#)]
234. Beheshti, I.; Mishra, S.; Sone, D.; Khanna, P.; Matsuda, H. T1-weighted mri-driven brain age estimation in Alzheimer’s disease and parkinson’s disease. *Aging Dis.* **2020**, *11*, 618. [[CrossRef](#)]
235. McCartney, D.L.; Stevenson, A.J.; Walker, R.M.; Gibson, J.; Morris, S.W.; Campbell, A.; Murray, A.D.; Whalley, H.C.; Porteous, D.J.; McIntosh, A.M.; et al. Investigating the relationship between dna methylation age acceleration and risk factors for Alzheimer’s disease. *Alzheimer’s Dement. Diagn. Assess. Dis. Monit.* **2018**, *10*, 429–437. [[CrossRef](#)] [[PubMed](#)]
236. Norrara, B.; Doerl, J.G.; Guzen, F.P.; Cavalcanti, J.R.L.P.; Freire, M.A.M. Commentary: Localized vs. systematic neurodegeneration: A paradigm shift in understanding neurodegenerative diseases. *Front. Syst. Neurosci.* **2017**, *11*, 91. [[CrossRef](#)] [[PubMed](#)]
237. Kim, Y.E.; Jung, Y.S.; Ock, M.; Yoon, S.J. Daly estimation approaches: Understanding and using the incidence-based approach and the prevalence-based approach. *J. Prev. Med. Public Health* **2022**, *55*, 10. [[CrossRef](#)]
238. Gao, T.; Wang, X.C.; Chen, R.; Ngo, H.H.; Guo, W. Disability adjusted life year (daly): A useful tool for quantitative assessment of environmental pollution. *Sci. Total Environ.* **2015**, *511*, 268–287. [[CrossRef](#)]
239. Jonsson, B.A.; Bjornsdottir, G.; Thorgeirsson, T.E.; Ellingsen, L.M.; Walters, G.B.; Gudbjartsson, D.F.; Stefansson, H.; Stefansson, K.; Ulfarsson, M.O. Brain age prediction using deep learning uncovers associated sequence variants. *Nat. Commun.* **2019**, *10*, 5409. [[CrossRef](#)]
240. Hung, C.W.; Chen, Y.C.; Hsieh, W.L.; Chiou, S.H.; Kao, C.L. Ageing and neurodegenerative diseases. *Ageing Res. Rev.* **2010**, *9*, S36–S46. [[CrossRef](#)]
241. Sibbett, R.A.; Altschul, D.M.; Marioni, R.E.; Deary, I.J.; Starr, J.M.; Russ, T.C. Dna methylation-based measures of accelerated biological ageing and the risk of dementia in the oldest-old: A study of the lothian birth cohort 1921. *BMC Psychiatry* **2020**, *20*, 1–15. [[CrossRef](#)]
242. Soldan, A.; Pettigrew, C.; Li, S.; Wang, M.C.; Moghekar, A.; Selnes, O.A.; Albert, M.; O’Brien, R. Relationship of cognitive reserve and cerebrospinal fluid biomarkers to the emergence of clinical symptoms in preclinical Alzheimer’s disease. *Neurobiol. Aging* **2013**, *34*, 2827–2834. [[CrossRef](#)]
243. Fries, G.R.; Bauer, I.E.; Scaini, G.; Valvassori, S.S.; Walss-Bass, C.; Soares, J.C.; Quevedo, J. Accelerated hippocampal biological aging in bipolar disorder. *Bipolar Disord.* **2020**, *22*, 498–507. [[CrossRef](#)] [[PubMed](#)]
244. Tønnesen, S.; Kaufmann, T.; de Lange, A.G.; Richard, G.; Doan, N.T.; Alnæs, D.; van der Meer, D.; Rokicki, J.; Moberget, T.; Maximov, I.I.; et al. Brain age prediction reveals aberrant brain white matter in schizophrenia and bipolar disorder: A multisample diffusion tensor imaging study. *Biol. Psychiatry Cogn. Neurosci. Neuroimaging* **2020**, *5*, 1095–1103. [[CrossRef](#)] [[PubMed](#)]
245. Heather, C.W.; Jude, G.; Riccardo, M.; Rosie, M.W.; Toni-Kim, C.; David, M.H.; Mark, J.A.; Lynsey, H.; Stewart, M.; Ian, J.D.; et al. Accelerated epigenetic ageing in major depressive disorder. *bioRxiv* **2017**. [[CrossRef](#)]
246. Dudley, J.A.; Lee, J.H.; Durling, M.; Strakowski, S.M.; Eliassen, J.C. Age-dependent decreases of high energy phosphates in cerebral gray matter of patients with bipolar i disorder: A preliminary phosphorus-31 magnetic resonance spectroscopic imaging study. *J. Affect. Disord.* **2015**, *175*, 251–255. [[CrossRef](#)] [[PubMed](#)]
247. Masuda, Y.; Okada, G.; Takamura, M.; Shibasaki, C.; Yoshino, A.; Yokoyama, S.; Ichikawa, N.; Okuhata, S.; Kobayashi, T.; Yamawaki, S.; et al. Age-related white matter changes revealed by a whole-brain fiber-tracking method in bipolar disorder compared to major depressive disorder and healthy controls. *Psychiatry Clin. Neurosci.* **2021**, *75*, 46–56. [[CrossRef](#)] [[PubMed](#)]
248. Muangpaisan, W.; Petcharat, C.; Srinonprasert, V. Prevalence of potentially reversible conditions in dementia and mild cognitive impairment in a geriatric clinic. *Geriatr. Gerontol. Int.* **2012**, *12*, 59–64. [[CrossRef](#)]
249. Gauthier, S.; Touchon, J. Mild cognitive impairment is not a clinical entity and should not be treated. *Arch. Neurol.* **2005**, *62*, 1164–1166. [[CrossRef](#)]
250. Bates, K.; Harvey, A.R.; Carruthers, M.; Martins, R. Androgens, andropause and neurodegeneration: Exploring the link between steroidogenesis, androgens and Alzheimer’s disease. *Cell Mol. Life Sci.* **2005**, *62*, 281–292. [[CrossRef](#)]

251. Liu, Z.; Zhou, T.; Ziegler, A.C.; Dimitrion, P.; Zuo, L. Oxidative stress in neurodegenerative diseases: From molecular mechanisms to clinical applications. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 2525967. [[CrossRef](#)]
252. Biswas, S.K. Does the interdependence between oxidative stress and inflammation explain the antioxidant paradox? *Oxidative Med. Cell. Longev.* **2016**, *2016*, 5698931. [[CrossRef](#)] [[PubMed](#)]
253. Bordoni, M.; Scarian, E.; Rey, F.; Gagliardi, S.; Carelli, S.; Pansarasa, O.; Cereda, C. Biomaterials in neurodegenerative disorders: A promising therapeutic approach. *Int. J. Mol. Sci.* **2020**, *21*, 3243. [[CrossRef](#)] [[PubMed](#)]
254. Bell, C.G.; Lowe, R.; Adams, P.D.; Baccarelli, A.A.; Beck, S.; Bell, J.T.; Christensen, B.C.; Gladyshev, V.N.; Heijmans, B.T.; Horvath, S.; et al. DNA methylation aging clocks: Challenges and recommendations. *Genome Biol.* **2019**, *20*, 1–24. [[CrossRef](#)] [[PubMed](#)]
255. Stubbs, T.M.; Bonder, M.J.; Stark, A.K.; Krueger, F.; von Meyenn, F.; Stegle, O.; Reik, W. Multi-tissue DNA methylation age predictor in mouse. *Genome Biol.* **2017**, *18*, 68. [[CrossRef](#)] [[PubMed](#)]
256. Thompson, M.J.; Chwiałkowska, K.; Rubbi, L.; Lusi, A.J.; Davis, R.C.; Srivastava, A.; Korstanje, R.; Churchill, G.A.; Horvath, S.; Pellegrini, M. A multi-tissue full lifespan epigenetic clock for mice. *Aging* **2018**, *10*, 2832. [[CrossRef](#)]
257. Wang, T.; Tsui, B.; Kreisberg, J.F.; Robertson, N.A.; Gross, A.M.; Yu, M.K.; Carter, H.; Brown-Borg, H.M.; Adams, P.D.; Ideker, T. Epigenetic aging signatures in mice livers are slowed by dwarfism, calorie restriction and rapamycin treatment. *Genome Biol.* **2017**, *18*, 57. [[CrossRef](#)]
258. Petkovich, D.A.; Podolskiy, D.I.; Lobanov, A.V.; Lee, S.G.; Miller, R.A.; Gladyshev, V.N. Using DNA methylation profiling to evaluate biological age and longevity interventions. *Cell Metab.* **2017**, *25*, 954–960. [[CrossRef](#)]
259. Meer, M.V.; Podolskiy, D.I.; Tyshkovskiy, A.; Gladyshev, V.N. A whole lifespan mouse multi-tissue DNA methylation clock. *eLife* **2018**, *7*, e40675. [[CrossRef](#)]
260. Bahar, R.; Hartmann, C.H.; Rodriguez, K.A.; Denny, A.D.; Busuttill, R.A.; Dollé, M.E.; Calder, R.B.; Chisholm, G.B.; Pollock, B.H.; Klein, C.A.; et al. Increased cell-to-cell variation in gene expression in ageing mouse heart. *Nature* **2006**, *441*, 1011–1014. [[CrossRef](#)]
261. Rimmelé, P.; Bigarella, C.L.; Liang, R.; Izac, B.; Dieguez-Gonzalez, R.; Barbet, G.; Donovan, M.; Brugnara, C.; Blander, J.M.; Sinclair, D.A.; et al. Aging-like phenotype and defective lineage specification in sirt1-deleted hematopoietic stem and progenitor cells. *Stem Cell Rep.* **2014**, *3*, 44–59. [[CrossRef](#)]
262. Cheung, P.; Vallania, F.; Warsinske, H.C.; Donato, M.; Schaffert, S.; Chang, S.E.; Dvorak, M.; Dekker, C.L.; Davis, M.M.; Utz, P.J.; et al. Single-cell chromatin modification profiling reveals increased epigenetic variations with aging. *Cell* **2018**, *173*, 1385–1397. [[CrossRef](#)] [[PubMed](#)]
263. Hernando-Herraez, I.; Evano, B.; Stubbs, T.; Commere, P.H.; Jan Bonder, M.; Clark, S.; Andrews, S.; Tajbakhsh, S.; Reik, W. Ageing affects DNA methylation drift and transcriptional cell-to-cell variability in mouse muscle stem cells. *Nat. Commun.* **2019**, *10*, 4361. [[CrossRef](#)] [[PubMed](#)]
264. Trapp, A.; Kerepesi, C.; Gladyshev, V.N. Profiling epigenetic age in single cells. *Nat. Aging* **2021**, *1*, 1189–1201. [[CrossRef](#)] [[PubMed](#)]
265. Hu, Y.; An, Q.; Guo, Y.; Zhong, J.; Fan, S.; Rao, P.; Liu, X.; Liu, Y.; Fan, G. Simultaneous profiling of mRNA transcriptome and DNA methylome from a single cell. *Single Cell Methods Seq. Proteom.* **2019**, *1979*, 363–377.
266. Angermueller, C.; Lee, H.J.; Reik, W.; Stegle, O. Deepcp: Accurate prediction of single-cell DNA methylation states using deep learning. *Genome Biol.* **2017**, *18*, 1–13.
267. Hamidouche, Z.; Rother, K.; Przybilla, J.; Krinner, A.; Clay, D.; Hopp, L.; Fabian, C.; Stolzing, A.; Binder, H.; Charbord, P.; et al. Bistable epigenetic states explain age-dependent decline in mesenchymal stem cell heterogeneity. *Stem Cells* **2017**, *35*, 694–704. [[CrossRef](#)]
268. de Lima Camillo, L.P.; Lapierre, L.R.; Singh, R. A pan-tissue DNA-methylation epigenetic clock based on deep learning. *npj Aging* **2022**, *8*, 4. [[CrossRef](#)]
269. Peleg, S.; Sananbenesi, F.; Zovoilis, A.; Burkhardt, S.; Bahari-Javan, S.; Agis-Balboa, R.C.; Cota, P.; Wittnam, J.L.; Gogol-Doering, A.; Opitz, L.; et al. Altered histone acetylation is associated with age-dependent memory impairment in mice. *Science* **2010**, *328*, 753–756. [[CrossRef](#)]
270. Stefanelli, G.; Azam, A.B.; Walters, B.J.; Brimble, M.A.; Gettens, C.P.; Bouchard-Cannon, P.; Cheng, H.M.; Davidoff, A.M.; Narkaj, K.; Day, J.J.; et al. Learning and age-related changes in genome-wide h2a.z binding in the mouse hippocampus. *Cell Rep.* **2018**, *22*, 1124–1131. [[CrossRef](#)]
271. Klein, H.U.; McCabe, C.; Gjonneska, E.; Sullivan, S.E.; Kaskow, B.J.; Tang, A.; Smith, R.V.; Xu, J.; Pfenning, A.R.; Bernstein, B.E.; et al. Epigenome-wide study uncovers tau pathology-driven changes of chromatin organization in the aging human brain. *Biorxiv* **2018**, *22*, 273789.
272. Roichman, A.; Elhanati, S.; Aon, M.A.; Abramovich, I.; Di Francesco, A.; Shahar, Y.; Avivi, M.Y.; Shurgi, M.; Rubinstein, A.; Wiesner, Y.; et al. Restoration of energy homeostasis by sirt6 extends healthy lifespan. *Nat. Commun.* **2021**, *12*, 3208. [[CrossRef](#)] [[PubMed](#)]
273. Grootaert, M.O.; Finigan, A.; Figg, N.L.; Uryga, A.K.; Bennett, M.R. Sirt6 protects smooth muscle cells from senescence and reduces atherosclerosis. *Circ. Res.* **2021**, *128*, 474–491. [[CrossRef](#)] [[PubMed](#)]
274. Soto-Palma, C.; Niedernhofer, L.J.; Faulk, C.D.; Dong, X. Epigenetics, DNA damage, and aging. *J. Clin. Investig.* **2022**, *132*, e158446. [[CrossRef](#)]
275. Hartmann, A.; Hartmann, C.; Secci, R.; Hermann, A.; Fuellen, G.; Walter, M. Ranking biomarkers of aging by citation profiling and effort scoring. *Front. Genet.* **2021**, *12*, 686320. [[CrossRef](#)]

276. Bürkle, A.; Moreno-Villanueva, M.; Bernhard, J.; Blasco, M.; Zondag, G.; Hoeijmakers, J.H.; Toussaint, O.; Grubeck-Loebenstein, B.; Mocchegiani, E.; Collino, S.; et al. MARK-AGE biomarkers of ageing. *Mech. Ageing Dev.* **2015**, *151*, 2–12. [[CrossRef](#)] [[PubMed](#)]
277. Mirzayans, R.; Murray, D. Human Genetic Disorders Associated with Genome Instability, Premature Aging and Cancer Predisposition. *Open Cancer J.* **2008**, *2*, 42–52. [[CrossRef](#)]
278. Tiwari, V.; Wilson, D.M. DNA damage and associated DNA repair defects in disease and premature aging. *Am. J. Hum. Genet.* **2019**, *105*, 237–257. [[CrossRef](#)]
279. Rizza, E.R.; DiGiovanna, J.J.; Khan, S.G.; Tamura, D.; Jeskey, J.D.; Kraemer, K.H. Xeroderma pigmentosum: A model for human premature aging. *J. Invest. Dermatol.* **2021**, *141*, 976–984. [[CrossRef](#)]
280. Han, Z.Z.; Fleet, A.; Larriou, D. Can accelerated ageing models inform us on age-related tauopathies? *Ageing Cell* **2023**, *22*, e13830. [[CrossRef](#)]
281. Platzer, M.; Englert, C. *Nothobranchius furzeri*: A model for aging research and more. *Trends Genet.* **2016**, *32*, 543–552. [[CrossRef](#)]
282. Reichwald, K.; Petzold, A.; Koch, P.; Downie, B.R.; Hartmann, N.; Pietsch, S.; Baumgart, M.; Chalopin, D.; Felder, M.; Bens, M.; et al. Insights into sex chromosome evolution and aging from the genome of a short-lived fish. *Cell* **2015**, *163*, 1527–1538. [[CrossRef](#)]
283. Valenzano, D.R.; Benayoun, B.A.; Singh, P.P.; Zhang, E.; Etter, P.D.; Hu, C.K.; Clément-Ziza, M.; Willemsen, D.; Cui, R.; Harel, I.; et al. The African turquoise killifish genome provides insights into evolution and genetic architecture of lifespan. *Cell* **2015**, *163*, 1539–1554. [[CrossRef](#)]
284. Petzold, A.; Reichwald, K.; Groth, M.; Taudien, S.; Hartmann, N.; Priebe, S.; Shagin, D.; Englert, C.; Platzer, M. The transcript catalogue of the short-lived fish *Nothobranchius furzeri* provides insights into age-dependent changes of mRNA levels. *BMC Genom.* **2013**, *14*, 185. [[CrossRef](#)] [[PubMed](#)]
285. Genade, T.; Benedetti, M.; Terzibasi, E.; Roncaglia, P.; Valenzano, D.R.; Cattaneo, A.; Cellerino, A. Annual fishes of the genus *Nothobranchius* as a model system for aging research. *Ageing Cell* **2005**, *4*, 223–233. [[CrossRef](#)] [[PubMed](#)]
286. Di Cicco, E.; Tozzini, E.T.; Rossi, G.; Cellerino, A. The shortlived annual fish *Nothobranchius furzeri* shows a typical teleost aging process reinforced by high incidence of age-dependent neoplasias. *Exp. Gerontol.* **2011**, *46*, 249–256. [[CrossRef](#)]
287. Dodzian, J.; Kean, S.; Seidel, J.; Valenzano, D.R. A protocol for laboratory housing of Turquoise Killifish (*Nothobranchius furzeri*). *J. Vis. Exp.* **2018**, *134*, e57073. [[CrossRef](#)]
288. Tan, L.; Ke, Z.; Tomblin, G.; Macoretta, N.; Hayes, K.; Tian, X.; Lv, R.; Ablaeva, J.; Gilbert, M.; Bhanu, N.V.; et al. Naked mole rat cells have a stable epigenome that resists iPSC reprogramming. *Stem Cell Rep.* **2017**, *9*, 1721–1734. [[CrossRef](#)]
289. Dammann, P.; Šumbera, R.; Massmann, C.; Scherag, A.; Burda, H. Extended longevity of reproductives appears to be common in *Fukomys molerats* (Rodentia, *Bathyergidae*). *PLoS ONE* **2011**, *6*, e18757. [[CrossRef](#)]
290. Ruby, J.G.; Smith, M.; Buffenstein, R. Naked mole-rat mortality rates defy gompertzian laws by not increasing with age. *Elife* **2018**, *7*, e31157. [[CrossRef](#)]
291. Munshi-South, J.; Wilkinson, G.S. Bats and birds: Exceptional longevity despite high metabolic rates. *Ageing Res. Rev.* **2010**, *9*, 12–19. [[CrossRef](#)]
292. Seim, I.; Fang, X.; Xiong, Z.; Lobanov, A.V.; Huang, Z.; Ma, S.; Feng, Y.; Turanov, A.A.; Zhu, Y.; Lenz, T.L.; et al. Genome analysis reveals insights into physiology and longevity of the Brandt’s bat *Myotis brandtii*. *Nat. Commun.* **2013**, *4*, 2212. [[CrossRef](#)] [[PubMed](#)]
293. Podlutsky, A.J.; Khritankov, A.M.; Ovodov, N.D.; Austad, S.N. A new field record for bat longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **2005**, *60*, 1366–1368. [[CrossRef](#)] [[PubMed](#)]
294. Holtze, S.; Lukač, M.; Cizelj, I.; Mutschmann, F.; Szentiks, C.A.; Jelić, D.; Hermes, R.; Göritz, F.; Braude, S.; Hildebrandt, T.B. Monitoring health and reproductive status of olms (*Proteus anguinus*) by ultrasound. *PLoS ONE* **2017**, *12*, e0182209. [[CrossRef](#)] [[PubMed](#)]
295. Mulec, J. Welcome to the -omics era of the 21st century: Will *Proteus anguinus* finally reveal all its mysteries? *Acta Carsol.* **2020**, *1*, 49. [[CrossRef](#)]
296. Voituron, Y.; de Fraipont, M.; Issartel, J.; Guillaume, O.; Clobert, J. Extreme lifespan of the human fish (*Proteus anguinus*): A challenge for ageing mechanisms. *Biol. Lett.* **2011**, *7*, 105–107. [[CrossRef](#)]
297. Philipp, E.E.; Wessels, W.; Gruber, H.; Strahl, J.; Wagner, A.E.; Ernst, I.M.; Rimbach, G.; Kraemer, L.; Schreiber, S.; Abele, D.; et al. Gene expression and physiological changes of different populations of the long-lived bivalve *Arctica islandica* under low oxygen conditions. *PLoS ONE* **2012**, *7*, e44621. [[CrossRef](#)]
298. Lutz, R.A.; Goodsell, J.G.; Mann, R.; Castagna, M. Experimental culture of the ocean quahog, *Arctica islandica*. *J. World Maricult. Soc.* **1981**, *12*, 196–205. [[CrossRef](#)]
299. Chapman, J.A.; Kirkness, E.F.; Simakov, O.; Hampson, S.E.; Mitros, T.; Weinmaier, T.; Rattei, T.; Balasubramanian, P.G.; Borman, J.; Busam, D.; et al. The dynamic genome of Hydra. *Nature* **2010**, *464*, 592–596. [[CrossRef](#)]
300. Schaible, R.; Sussman, M.; Kramer, B.H. Aging and potential for self-renewal: Hydra living in the age of aging—A mini-review. *Gerontology* **2014**, *60*, 548–556. [[CrossRef](#)]
301. Bellantuono, A.J.; Bridge, D.; Martínez, D.E. Hydra as a tractable, long-lived model system for senescence. *Invert. Reprod. Dev.* **2015**, *59*, 39–44. [[CrossRef](#)]
302. Klimovich, A.; Rehm, A.; Wittlieb, J.; Herbst, E.-M.; Benavente, R.; Bosch, T.C.G. Non-senescent Hydra tolerates severe disturbances in the nuclear lamina. *Ageing* **2018**, *10*, 951–972. [[CrossRef](#)] [[PubMed](#)]

303. Grohme, M.A.; Schloissnig, S.; Rozanski, A.; Pippel, M.; Young, G.R.; Winkler, S.; Brandl, H.; Henry, I.; Dahl, A.; Powell, S.; et al. The genome of *Schmidtea mediterranea* and the evolution of core cellular mechanisms. *Nature* **2018**, *554*, 56–61. [[CrossRef](#)] [[PubMed](#)]
304. Fincher, C.T.; Wurtzel, O.; de Hoog, T.; Kravarik, K.M.; Reddien, P.W. Cell type transcriptome atlas for the planarian *Schmidtea mediterranea*. *Science* **2018**, *360*, eaaq1736. [[CrossRef](#)]
305. Merryman, M.S.; Alvarado, A.S.; Jenkin, J.C. Culturing Planarians in the Laboratory. *Methods Mol. Biol.* **2018**, *1774*, 241–258. [[CrossRef](#)]
306. Lipsky, M.S.; King, M. Biological theories of aging. *Dis.-A-Mon.* **2015**, *61*, 460–466. [[CrossRef](#)] [[PubMed](#)]
307. Jin, K. Modern biological theories of aging. *Aging Dis.* **2010**, *1*, 72. [[PubMed](#)]
308. Weinert, B.T.; Timiras, P.S. Invited review: Theories of aging. *J. Appl. Physiol.* **2003**, *95*, 1706–1716. [[CrossRef](#)] [[PubMed](#)]
309. Northrop, J.H. The influence of the intensity of light on the rate of growth and duration of life of *Drosophila*. *J. Gen. Physiol.* **1925**, *9*, 81. [[CrossRef](#)]
310. Northrop, J.H. Duration of life of an aseptic *Drosophila* culture inbred in the dark for 230 generations. *J. Gen. Physiol.* **1926**, *9*, 763–765. [[CrossRef](#)]
311. NCBI PubMed. Available online: <https://pubmed.ncbi.nlm.nih.gov/> (accessed on 31 August 2023).
312. Tilney, F. The Aging of the Human Brain. *Bull. N. Y. Acad. Med.* **1928**, *4*, 1125–1143.

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.